

UNIVERSITY OF KERALA

THIRUVANANTHAPURAM



COURSE STRUCTURE AND SYLLABUS

For

FIRST DEGREE PROGRAMME

IN

BOTANY

UNDER

CHOICE BASED CREDIT- SEMESTER SYSTEM

(w.e.f. 2019 admission onwards)

OBJECTIVES OF THE PROGRAMME

- ✎ To impart knowledge of Science is the basic objective of education.
- ✎ To develop scientific attitude is the major objective to make the students open minded, critical,curious.
- ✎ To develop skill in practical work, experiments and laboratory materials and equipments along with the collection and interpretation of scientific data to contribute the science.
- ✎ To understand scientific terms, concepts, facts, phenomenon and their relationships.
- ✎ To make the students aware of natural resources and environment.
- ✎ To provide practical experience to the students as a part of the course to develop scientific ability to work in the field of research and other fields of their own interest and to make them fit for society.
- ✎ The students are expected to acquire knowledge of plant and related subjects so as to understand natural phenomenon, manipulation of nature and environment for the benefit of human beings.
- ✎ To develop ability for the application of the acquired knowledge to improve agriculture and other related fields to make the country self reliant and sufficient.
- ✎ Understand and appreciate the role of biology in societal issues, such as the environment and biological resources, biodiversity, ethics and human health and diseases.
- ✎ To enrich the students with the latest developments in the field of Information technology, Biotechnology, Bio informatics and other related fields of research and development.
- ✎ To create enthusiasm to understand more about the beautiful planet Earth and to give awareness to the public the need to protect the planet from all kinds of exploitation.
- ✎ To keep the scientific temper which the student acquired from school level and to develop a research culture.

Table 1. General Structure of the First Degree Programme in Botany

Sem No.	Course Code	Course Title	Instructional hours/ week		Credits	Uty Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
I	EN1111	English Language I Additional	5	-	4	3hrs	20%	80%	16
	1111	Additional Language I	4	-	3	„			
	EN1121	Foundation Course I	4	-	2	„			
	BO1141	Core Course I	2	2	3	„			
	CH/BC1131	Compl. Course I (CH/BC)	2	2	2	„			
	ZO1131	Complementary Course II (ZO)	2	2	2	„			
II	EN1211	English Language II	5	-	4	„	20%	80%	17
	EN1212	English Language III	4	-	3	„			
	1211	Additional Language II	4	-	3	„			
	BO1221	Foundation Course II	2	2	3	„			
	CH/BC1231	Compl. Course III(CH/BC)	2	2	2	„			
	ZO1231	Compl. Course IV(ZO)	2	2	2	„			
III	EN1311	English Language IV	5	-	4	„	20%	80%	17
	1311	Additional Language III	5	-	4	„			
	BO1341	Core Course II	3	2	3	„			
	CH/BC1331	Compl. Course V(CH/BC)	3	2	3	„			
	ZO1331	Compl. Course VI(ZO)	3	2	3	„			
IV	EN1411	English Language V	5	--	4	„	20%	80%	25
	1411	Additional Language IV	5	-	4	„			
	BO1441	Core Course III	3	2	3	„			
	CH/BC1431	Compl.CourseVII(CH/BC)	3	2	3	„			
	ZO1431	Compl. courseVIII(ZO)	3	2	3	„			
	CH/BC1432	Compl IX (Practical CH/BC)	-	(8)*	4	„			
	ZO1432	Compl X (Practical ZO)	-	(8)*	4	„			
V	BO1541	Core Course IV	4	3	4	„	20%	80%	20
	BO1542	Core Course V	5	2	4	„			
	BO1543	Core Course VI	4	2	3	„			
	BO1544	Core (Practical-1) VII	-	(4)*	3	„			
	BO1545	Core (Practical-2)VIII	-	(4)*	4	„			
		Open Course I	3	-	2	„			
		BO1551.1 Horticulture							
		BO1551.2 Mushroom cultivation & Marketing							
		BO1551.3 Forestry							
		Project	-	2					
VI	BO1641	Core Course IX	5	2	4	„	20%	80%	25
	BO1642	Core Course X	4	2	4	„			
	BO1643	Core Course XI	4	2	4	„			
	BO1644	Core (Practical-3)XII	-	(5)*	3	„			
	BO1645	Core (Practical-4)XIII	-	(8)*	4	„			
	BO1651	Elective Course II	3	-	2	„			
	BO1646	Project		3	4				
								120	

L = Lecture P = Practical ()*Practical hour already distributed in the semester concerned

Table 2. SEMESTER – I

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
I	EN1111	English Language I	5	-	4	3hrs	20%	80%	16
	1111	Additional Language I	4	-	3	„			
	EN1121	Foundation Course I	4	-	2	„			
	BO1141	Core Course I -Angiosperm Anatomy Reproductive Botany & Palynology	2	2	3	„			
	CH/BC1131	Compl. Course I (CH/BC)	2	2	2	„			
	ZO1131	Compl. Course II(ZO)	2	2	2	„			

Table 3. SEMESTER – II

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
II	EN1211	English Language II	5	-	4	3hrs	20%	80%	17
	EN1212	English Language III	4	-	3	„			
	1211	Additional Language II	4	-	3	„			
	BO1221	Foundation Course II- Methodology & Perspectives in Plant Science	2	2	3	„			
	CH/BC1231	Compl. Course III(CH/BC)	2	2	2	„			
	ZO1231	Compl. Course IV(ZO)	2	2	2	„			

Table 4. SEMESTER – III

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty. Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
III	EN1311	English Language IV	5	-	4	3hrs	20%	80%	17
	1311	Additional Language III Core	5	-	4	„			
	BO1341	Course II- Microbiology, Phycology, Mycology, Lichenology & Plant Pathology	3	2	3	„			
	CH/BC1331	Compl. Course V(CH/BC)	3	2	3	„			
	ZO1331	Compl. Course VI(ZO)	3	2	3	„			

Table 5. SEMESTER –IV

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty. Exam Duration	Evaluation		Total credit
			T	P			Internal	University exam	
IV	EN1411	English Language V	5	-	4	3 hrs	20%	80%	25
	1411	Additional Language IV	5	-	4	„			
	BO1441	Core Course III- Bryology, Pteridology, Gymnosperms & Paleobotany	3	2	3	„			
	CH/BC1431	Compl. Course VII(CH/BC)	3	2	3	„			
	ZO1431	Complementary Course VIII (ZO)	3	2	3	„			
	CH/BC1432	Compl IX (Practical CH/BC)	-	(8)*	4	„			
	ZO1432	Compl X (Practical ZO)	-	(8)*	4	„			

Table 6. SEMESTER - V

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty. Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
V	BO1541	Core Course IV - Angiosperm Morphology, Systematic botany, Economic botany, Ethno botany & Pharmacognosy	4	3	4	3hrs	20%	80%	20
	BO1542	Core Course V- Environmental Studies & Phytogeography	5	2	4	„			
	BO1543	Core Course VI- Cell Biology, Genetics & Evolutionary Biology	4	2	3	„			
	BO1544	Core (Practical-1) VII (BO1141, BO1221)	-	(4)*	3	„			
	BO1545	Core (Practical-2) VIII (BO1341 & BO1441)	-	(4)*	4	„			
		Open Course I	3	-	2	„			
	BO1551.1	Horticulture							
	BO1551.2	Mushroom cultivation							
		& Marketing							
	BO1551.3	Forestry							
		Project	-	2	-				

()*Practical hour already distributed in the semester concerned

Table 7. SEMESTER – VI

Sem No.	Course Code	Course Title	Instructional hours/week		Credits	Uty. Exam Duration	Evaluation		Total credits
			T	P			Internal	University exam	
VI	BO1641	Core Course IX Plant physiology &Biochemistry	5	2	4	3hrs	20%	80%	25
	BO1642	Core Course X- Molecular Biology, General informatics and Bioinformatics	4	2	4	„			
	BO1643	Core Course XI- Plant Breeding, Horticulture& Research methodology	4	2	4	“			
	BO1644	Core (Practical-3)XII (BO1541 & BO1542)	-	(5)*	3	„			
	BO1645	Core (Practical-4) XIII IV (BO1543, BO1641, BO1642, 1643)	-	(8)*	4	„			
	BO1651	Open Course II- Elective- Biotechnology & Nano Biotechnology	3	-	2	„			
	BO1646	Project	-	3	4				

Table 8. Distribution of Contact Hours and Credits
(CORE, FOUNDATION & OPEN COURSES, PROJECT/DISSERTATION)

Course Code	Course Title	Semester I			Semester II			Semester III			Semester IV			Semester V			Semester VI			Total	
		Contact hours		Credit	Contact hours		Credit	Contact hours		Credit	Contact hours		Credit	Contact hours		Credit	Contact hours		Credit	Contact hours	Credit
		T	P		T	P		T	P		T	P		T	P		T	P			
BO1141	Angiosperm anatomy, Reproductive Botany & Palynology	2	2	3															4	3	
BO1221	Methodology & Perspectives in Plant Science				2	2	3												4	3	
BO1341	Microbiology, Phycology, Mycology Lichenology & Plant Pathology							3	2	3									5	3	
BO1441	Bryology, Pteridology, Gymnosperms & Paleobotany										3	2	3						5	3	
BO1541	Angiosperm Morphology Systematic botany, Economic botany Ethno botany & Pharmacognosy													4	3	4			7	4	
BO1542	Environmental Studies & Phytogeography													5	2	4			7	4	
BO1543	Cell Biology, Genetics & Evolutionary Biology													4	2	3			6	3	
BO1544	Practical-I (BO1141, BO1221)		2*		2*														4	3	
BO1545	Practical-II (BO1341 & BO1441)							*2			2*								4	4	
BO1551.1 BO1551.2 BO1551.3	Horticulture, Mushroom cultivation & Marketing Forestry													3					3	2	
BO1641	Plant Physiology & Biochemistry																5	2	4	7	4
BO1642	Molecular Biology, General informatics and Bioinformatics																4	2	4	6	4
BO1643	Plant breeding, Horticulture& Research methodology																4	2	4	6	4
BO1644	Practical-III (BO1541 & BO1542)													5*							3
BO1645	Practical-IV (BO1543, BO1641, BO1642, 1643)																	8*			4
BO1651	Biotechnology & Nano biotechnology																3		3	2	
BO1646	Project report, Tour Diary, Viva-Voce													2			3		5	4	
	Total																				57

L = Lecture P = Practical *Practical hour already distributed in the semester concerned

Table 9. Scheme of Evaluation of Foundation Course II, Core Courses, Open Courses & Project

Semester	Course Code	Course Title	Marks		Duration of University Exam.
			CE	ESE	
I	BO1141	Angiosperm anatomy, Reproductive Botany & Palynology	20	80	3 hrs
II	BO1221	Methodology & Perspectives in Plant Science	20	80	3 hrs
III	BO1341	Microbiology, Phycology, Mycology, Lichenology & Plant Pathology	20	80	3 hrs
IV	BO1441	Bryology, Pteridology, Gymnosperms & Paleobotany	20	80	3 hrs
V	BO1541	Angiosperm Morphology, Systematic botany, Economic botany, Ethnobotany & Pharmacognosy	20	80	3 hrs
	BO1542	Environmental Studies & Phytogeography	20	80	3 hrs
	BO1543	Cell biology, Genetics & Evolutionary Biology	20	80	3 hrs
	BO1544	Practical-I (BO1141, BO1221)	20	80	3 hrs
	BO1545	Practical-II (BO1341 & BO1441)	20	80	3 hrs
	BO1551.1 BO1551.2 BO1551.3	Horticulture, Mushroom cultivation & Marketing, Forestry	20	80	3 hrs
VI	BO1641	Plant Physiology & Biochemistry	20	80	3 hrs
	BO1642	Molecular Biology, General informatics and Bioinformatics	20	80	3 hrs
	BO1643	Plant breeding, Horticulture & Research Methodology	20	80	3 hrs
	BO1644	Practical-III (BO1541 & BO1542)	20	80	3 hrs
	BO1645	Practical-IV (BO1543, BO1641, BO1642, 1643)	20	80	3 hrs
	BO1651	Biotechnology & Nano biotechnology	20	80	3 hrs
	BO1646	Project report, Tour Diary, Viva-Voce	20	80	-

END SEMESTER ASSESSMENT (ESA)

The University shall conduct the external examinations for all semesters. There will not be any supplementary exams. The practical examinations for **Core courses** shall be conducted after 4th, 5th and 6th semesters and **Complementary** courses at the end of 4th semester according to the common calendar and questions set up by the University. The Board of Examiners constituted by the University will have the right to make necessary changes in the pattern of practical examination as and when needed with the prior sanction of the Chairman, Board of Studies Botany (Pass). The practicals mentioned in the syllabus under various branches of Botany Core and Complementary Courses should be incorporated in the Practical record.

ELIGIBILITY TO APPEAR FOR PRACTICAL EXAMINATION

Submission of the following

- ✗ Certified and bonafide practical record
- ✗ Certified herbarium sheets
- ✗ Certified field Book
- ✗ Certified Tour Report
- ✗ Project report/Dissertation (certified and bonafide)

PROJECT

Project work/Dissertation is compulsory. Students have to begin the project in the 5th Semester and submit the project report for valuation at the end of 6th Semester. Viva-Voce may be conducted for each student at the time of Project evaluation. Project can be carried out either individually or by a group not exceeding 15 students ensuring direct and active participation of each student in the group. The topics shall either be allotted by the supervising teacher or be selected by the student in consultation with the supervising teacher. The project report/dissertation duly attested by the Supervising teacher and Certified by the Head of the Department, has to be submitted on the day of examination of **Practical - III (Core)**. The project shall be evaluated by an external examiner. The project report/ Dissertation (not less than 30 pages) shall be prepared in Times New Roman font size 12 with 1.5 spacing as per the format given below.

1. Title page /Front page (Certified by the **HOD**)
2. Declaration by the candidate
3. Certificate attested by the Supervising Teacher
4. Acknowledgement, if any
5. Table of contents
6. Abbreviation, if any
7. Introduction & Review of Literature
8. Material and Methods
9. Results and Discussion (Not less than 10 pages)
10. Summary and Conclusion
11. References

Tables, Graphs, Photographs etc. can be used to present the data. Topics selected once should not be repeated and plagiarism should be avoided.

STUDY TOUR

- ✎ Field trip to a place of plant diversity within or outside Kerala with a minimum duration of 3 days is compulsory. (Field trips are to be conducted for three days either as continuous or one day trips).
- ✎ A brief report of the trip has to be submitted at the time of Practical Examination

CORE COURSES

Semester	Course Code	Course Title	Contact hrs/ week		Credits
			L	P	
I	BO1141	Angiosperm anatomy Reproductive Botany & Palynology	2	2	3
III	BO1341	Microbiology, Phycology, Mycology Lichenology & Plant Pathology	3	2	3
IV	BO1441	Bryology, Pteridology, Gymnosperms & Paleobotany	3	2	3
V	BO1541	Angiosperm Morphology Systematic botany, Economic botany Ethno botany & Pharmacognosy	4	3	4
	BO1542	Environmental Studies & Phytogeography	5	2	4
	BO1543	Cell Biology, Genetics & Evolutionary Biology	4	2	3
	BO1544	Practical-I (BO1141, BO1221)	-	4	3
	BO1545	Practical-II (BO1341 & BO1441)	-	4	4
VI	BO1641	Plant Physiology & Biochemistry	5	2	4
	BO1642	Molecular Biology, General informatics and Bioinformatics	4	2	4
	BO1643	Plant breeding, Horticulture & Research methodology	4	2	4
	BO1644	Practical-III (BO1541 & BO1542)	-	5	3
	BO1645	Practical-IV (BO 1543, BO1642, BO1642 & BO1643)	-	8	4

SEMESTER-I

ANGIOSPERM ANATOMY, REPRODUCTIVE BOTANY AND PALYNOLOGY

Course code BO 1141, Number of credits: 3

Number of contact hours: Lecture: 36 hrs; Practical: 36 hrs; Total: 72 hrs

Distribution of Hours	Theory	Practical
Angiosperm Anatomy	27 hrs	27 hrs
Reproductive Botany & Palynology	09 hrs	09 hrs
Total	36 hrs	36 hrs

Aim of the course: To generate awareness about anatomical features of Angiosperms & Reproductive biology.

Objectives:

- ✎ To develop skills for identification of microscopic structures
- ✎ To distinguish various tissue systems and internal structure
- ✎ To acquire basic knowledge about embryo development and pollen grains

ANGIOSPERM ANATOMY

MODULE – 1

04 hrs

1. Objective and scope of plant anatomy
2. Cell wall organisation – Gross structure- Primary and secondary wall, pits-plasmodesmata-microscopic and submicroscopic structures- Extra cell wall material (lignin, suberin), Non living inclusions of the cell [Reserve food (carbohydrates, proteins, fats and oil), secretory products (colouring matter, enzymes, nectar), excretory products (nitrogenous and non nitrogenous including resins, tannins, organic acids, latex, essential oils, glycosides, gums and mineral crystals)].

MODULE - II

11 hrs

1. Tissues-Meristems: Definition, Classification based on origin, position, growth patterns, functions
2. Apical meristem & theories on apical organisation- Apical cell theory, Histogen theory, Tunica-Corpus theory. Organisation of shoot apex in dicots and monocots (Korper Kappe theory)
3. Permanent tissues – Definition, classification – simple, complex and secretory tissues (glandular tissue, laticifers).
4. Tissue systems- Epidermal tissue system, Ground tissue system and vascular tissue systems. Stomata – structure and functions, types- anomocytic, anisocytic, paracytic, diacytic, graminaceous. Different types of vascular arrangements.

SEMESTER-I

MODULE- III

12 hrs

1. Primary structure – Root, stem and leaf (Dicot & Monocot)
2. Secondary growth- Root and stem- cambium (structure and function), annual rings, heart wood and sap wood, hard wood and soft wood, tyloses, ring porous wood and diffuse porous wood, periderm formation – phellum, phellogen and phelloderm; lenticels
3. Anomalous secondary growth – *Bignonia*, *Boerhaavia*, *Dracaena*.

Practicals

27 hrs

1. Non-living inclusions – Cystolith, Raphide, Sphaero- raphide (Druses), Aleurone grains
2. Starch grains (Eccentric, Concentric, compound)
3. Simple permanent tissue- Parenchyma, Chlorenchyma, Aerenchyma, Collenchyma, Sclerenchyma
4. Primary structure- Dicot stem- *Centella*, *Chromolaena*
5. Monocot stem- Grass and *Asparagus*
6. Dicot root- Pea/ *Limnanthemum*
7. Monocot root- Colocasia or any other monocot root
8. Secondary structure – Stem (Normal type) – *Vernonia*
9. Secondary structure – Root (Normal type)- *Carica papaya*, Aerial root- *Tinospora* and *Ficus*
10. Epidermal structures- Stomata (Anomocytic, anisocytic, paracytic, diacytic)
11. Anomalous secondary thickening – *Bignonia*, *Boerhaavia*, *Dracaena*

REPRODUCTIVE BOTANY AND PALYNOLOGY

MODULE – IV

09 hrs

1. Introduction to angiosperm embryology, Contribution of Indian embryologists
2. P. Maheswary
3. Microsporogenesis- Structure and functions of wall layers
4. Development of male gametophyte- Dehiscence of anther
5. Megasporogenesis – Development of female gametophyte – Embryo sac- Development and types – Monosporic – *Polygonum* type, Bisporic – *Allium* type, Tetrasporic – *Adoxa* type
6. Pollination- Germination of pollen grains, Fertilization, double fertilization, Barriers of fertilization
7. Structure of embryo – Dicot (*Capsella*). Monocot (*Sagittaria*), Endosperm types (Cellular, nuclear and helobial – Brief account only)
8. Palynology: Pollen structure, aperture morphology, pollen allergy. Economic and taxonomic importance (Brief account only)

Practical

09 hrs

1. Pollen morphoforms (Colpate, porate and colporate- Photographs/permanent slides. SEM images not needed)
2. Students should be familiar with the structure of anther and embryo (Permanent slides can be used)

SEMESTER-I

REFERENCES

1. Bhattacharya K and Majumdar M R.(2011) A text book of palynology, New Central Book Agency, Calcutta
2. Bhojwani S S, Dantu P K & Bhatnagar S P (2014) The embryology of Angiosperms, Vikas Publishing
3. Coutler E. G. (1969) Plant Anatomy – Part I Cells and Tissues – Edward Arnold, London
4. David F. Cutler , Ted Botha and Dennis W M. Stevenson (2008) Plant Anatomy: An Applied Approach, John Wiley and Sons Ltd
5. Esau K (1965) Plant Anatomy- Wiley Eastern, New York
6. Esau K (2006) Anatomy of seed plants 2nd edition Wiley Eastern, New York
7. Fahn A (1995) – Plant Anatomy , Elsevier Science and Technology, Oxford, UK
8. Johri B M (2011) Embryology of Angiosperms, Springer
9. Kashinath Bhattacharya, Manas Ranjan Majumdar, Swati Gupta Bhattacharya (2011) A Text Book of Palynology, New Central Book Agency; 3rd Revised edition
10. Maheswari P (2011) An Introduction to the Embryology of Angiosperms, McGraw Hill, New York
11. Nair P K K (1970) Palynology of Angiosperms. Vikas Publishing House, New Delhi
12. Pandey S N and Chadha A (1997)– Plant Anatomy and Embryology, Vikas Publishing
13. Pandey AK (2000) Introduction to Embryology of Angiosperms, CBS Publishers & Distributors
14. Pandey B P (1997) Plant Anatomy – S Chand and Co. New Delhi Biology- Mc Graw Hill Co, New York
15. Pandey B P (2012) – Plant Anatomy. S Chand Publishing
16. Pijush Roy (2006) Plant Anatomy. New Central Book Agency (P) Ltd
17. Vashista P C (1984) – Plant Anatomy, Pradeep Publications, Jalandhar

EXPECTED OUTCOME

- ✎ Students are able to understand the complexities of cell wall organization, microscopic and sub microscopic structures.
- ✎ Students can distinguish various anatomical features of monocots and dicots (stem and root) with respect to permanent tissues and tissue systems.
- ✎ Identify and differentiate male and female gametophyte development in angiosperms.
- ✎ Distinguish monocot and dicot embryo and the basic features of pollen grains.

SEMESTER-II

FOUNDATION COURSE

METHODOLOGY AND PERSPECTIVES IN PLANT SCIENCES

Course code : BO 1221, Number of credits : 3

Number of contact hours: 36 hrs (Lecture); 36hrs (Practical) Total= 72 hours

Distribution of Hours	Theory	Practical
Scientific Studies	08 hrs	0 hrs
Data handling in science	12 hrs	16 hrs
Microtechnique	06 hrs	08 hrs
Biophysics	10 hrs	12 hrs
Total	36 hrs	36 hrs

Aim of the course: To introduce the methodology and perspectives of Science in general so as to enable the students to systematically pursue his particular discipline in science in relation to other disciplines that come under the rubric of sciences.

Objectives:

- ☒ To familiarize the students with the fundamental characteristics of science and significance of scientific studies
- ☒ To apply scientific methods independently and familiarize instruments in biological labs
- ☒ To interpret scientific data using basic statistical methods
- ☒ To develop skills for microscopic specimen preparation.

MODULE-I

Scientific Studies

08hrs

1. Types of knowledge: Practical, theoretical and scientific knowledge
2. Information: What is science; what is not science; laws of science; basis of scientific laws and factual truths.
3. Science as a human activity: scientific temper and empiricism, Science disciplines
4. Revolution in Science and Technology
5. Ethics in Science: Transparency and Honesty; danger of pre conceived ideas.
6. Scientific information: Depositories of scientific information, Primary, secondary and digital sources; sharing of knowledge.
7. Methods and tools in science: Steps in scientific method. Null hypothesis and alternative hypothesis. Inductive and deductive reasoning.

MODULE- II

Data handling in science

12hrs

1. Nature and types of data - Typical examples, Data collection, Data presentation-

SEMESTER-II

Classification and tabulation, diagrammatic presentation (bar & pie diagrams) and graphic presentation (Histogram, frequency polygon, frequency curve & Ogives).

2. Samples and sampling techniques.
3. Statistical treatment of data: Statistical terms and symbols. Measures of central tendencies (mean, median, mode), Measures of dispersion (range, mean deviation, variance, standard deviation, coefficient of variation), Significance tests (chi-square test).

Practicals

16hrs

1. Workout problems on frequency distribution
2. Represent data using bar diagram and histogram
3. Measures of central tendencies (Mean, Median, Mode)
4. Workout problems on measures of dispersion (range, mean deviation, variance, standard deviation, coefficient of variation)
5. Workout problems on chi-square test.

MODULE-I11

Microtechnique

06hrs

1. Introduction - microscopy - simple and compound – phase contrast; dark field illumination and electron microscopes (SEM and TEM).
2. Micrometry, Camera lucida
3. Sectioning - hand and microtome – rotary and sledge
4. Killing and fixation agents – Carnoy's formula, Farmers formula, F .A.A
5. Dehydration - reagents
6. Stains and staining techniques - double staining. General account; Stains: safranin, haematoxylin, acetocarmine.
7. Mounting media - D. P. X and Canadabalsam
8. Whole mounts - cytological methods: maceration, smear and squash preparation.

Practical

08hrs

1. Familiarize stains, fixatives and mounting media
2. General awareness of Micro technique - maceration, smears & squash
3. Demonstration of microtome sectioning and hand sectioning
4. Measurement of specimens using micrometer (Demonstration only).
5. Photomicrography and Camera lucida drawings (Demonstration only).

MODULE-IV

Biophysics

10hrs

1. Principles and applications of Colorimeter, Spectrophotometer (UV-Visible) and Centrifuge- centrifugation, types of rotors (swinging bucket, fixed angle), Density gradient and Differential centrifugation.
2. Basic knowledge of the separation methods: - Chromatography (Column chromatography, paper chromatography and TLC), Electrophoresis (PAGE and AGE).
3. Buffers -their functions in biological systems - Uses of buffers in biological research, pH meter.
4. Cryobiology – cryopreservation, freeze drying (lyophilisation) and its applications.

SEMESTER-II

Practical

12hrs

1. Separation of plant pigments by paper chromatography/TLC.
2. Preparation of buffer
3. Measurement of pH
4. Construct the absorption spectrum of any sample.

REFERENCE

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EXPECTED OUTCOME

- ✘ Students will be familiarized with the fundamental characteristics of Science.
- ✘ Develops an idea about involvement of science in improvement of human life.
- ✘ Create awareness of scientific approach towards life and learns the values of ethics in science.
- ✘ Develops skills to interpret scientific data using basic statistical methods.
- ✘ Create skills to prepare specimens for microscopic and gross anatomical studies and familiarize with different microscopic methods for sample analysis.
- ✘ Students become able to prepare buffers, measure pH, separate plant pigments and construct absorption spectrum of a sample.

SEMESTER-III

MICROBIOLOGY, PHYCOLOGY, MYCOLOGY, LICHENOLOGY AND PLANT PATHOLOGY

Course code : BO 1341, Number of credits : 3

Number of contact hours: Lecture: 54 hrs; Practical: 36 hrs; Total: 90 hrs

Distribution of Hours	Theory	Practical
Microbiology	08 hrs	07 hrs
Phycology	20hrs	15 hrs
Mycology & Lichenology	20 hrs	10 hrs
Plant Pathology	06 hrs	04 hrs
Total	54 hrs	36 hrs

Aim of the course: To create awareness about the world of microbes and lower groups of plants.

Objectives:

- ✎ To familiarize characteristic features of microbes and their significance
- ✎ To create awareness about importance of microbes in environment
- ✎ To generate idea about types of algae, fungi, lichen and their economic as well as evolutionary significance

MODULE-I

Microbiology

08hrs

1. History & scope of microbiology.
2. Bacterial classification: Morphological classification, classification based on staining reaction: Ultra structure of bacteria, Reproduction, Economic importance.
3. Mycoplasma & Actinomycetes –General account.
4. Virus-General characteristics, Nomenclature, classification, structure, Chemical composition, properties and reproduction of bacteriophages and T. M. V.

Economic importance

5. Soil microbiology – Soil microorganisms, the rhizosphere
6. Aquatic microbiology - Microbiology of sewage or waste water. Methods of waste water treatment (Brief account only)
7. Food microbiology - Food spoilage and preservation methods [General account].
8. Agricultural microbiology - Role of microbes in soil fertility, Nitrogen fixation, Biofertilizers

Practical

07hrs

1. Gram staining of bacteria.
2. Test for the Coli form bacteria in contaminated water.

SEMESTER-III

3. Isolation of *Rhizobium* from root nodules of leguminous plants (Demonstration)
4. Examination of different forms of bacteria (Demonstration)

MODULE - II

Phycology

20hrs

1. Introduction – Range of thallus structure – Phylogenetic trends – Pigments – Reproduction
2. Life cycle – Classification proposed by F.E. Fritsch
3. Salient features of the following major groups with reference to the structure, reproduction and life cycle of the types given below **(Excluding the developmental details)**
 - a. Cyanophyceae – *Nostoc*
 - b. Chlorophyceae - *Chlorella*, *Volvox*, *Oedogonium* and *Chara*
 - c. Xanthophyceae – *Vaucheria*
 - d. Bacillariophyceae – *Pinnularia*
 - e. Phaeophyceae – *Sargassum*
 - f. Rhodophyceae – *Polysiphonia*

Economic importance of algae

- a. Commercial products of algae – Agar, Alginates, Carrageenin, Diatomaceous earth
- b. Algae - medicinal aspects, algal blooms and red tide

Practical

15hrs

1. Make micro preparations of vegetative and reproductive structures of the types mentioned in the syllabus.
2. Identify the algal specimens up to the generic level and make labeled sketches of the specimens observed.

MODULE-III

Mycology & Lichenology

20hrs

1. Introduction, structure, reproduction, life cycle, evolutionary trends, Classification based on Ainsworth (1972)
2. Distinguishing characters of different classes of fungi representing the following genera (Excluding Developmental details)
 - a. Myxomycotina – General characters.
 - b. Zygomycotina - *Rhizopus*
 - c. Ascomycotina
 - ☒ Hemiascomycetes - *Saccharomyces*
 - ☒ Plectomycetes - *Penicillium*
 - ☒ Pyrenomycetes - *Xylaria*
 - ☒ Discomycetes – *Peziza*
 - d. Basidiomycotina
 - ☒ Teliomycetes - *Puccinia*
 - ☒ Hymenomycetes - *Agaricus*
 - e. Deuteromycotina - *Cercospora*.

SEMESTER-III

3. Economic importance of Fungi

Lichenology: General account and economic importance; the structure, reproduction and life cycle of *Usnea*

Practicals

10hrs

- ✎ A detailed study of structure and reproductive structures of types given in the syllabus and submission of record
- ✎ *Rhizopus*, *Saccharomyces*, *Penicillium*, *Xylaria*, *Peziza*, *Puccinia*, *Agaricus*, *Cercospora* and *Usnea*.

MODULE-IV

Plant Pathology

06hrs

1. Classification of plant diseases on the basis of causative organisms and symptoms – Host-parasite interaction, phytoalexins.
2. Study of the following diseases with emphasis on symptoms, disease cycle and control measures - Leaf mosaic of Tapioca, Citrus Canker, Blast disease of Paddy, Root wilt of Coconut
3. Brief account of the following fungicides- Bordeaux mixture, Lime sulphur, Tobacco decoction, Neem cake & oil.

Practical

04hrs

1. Identify the Diseases mentioned with respect to causal organism and symptoms- Leafmosaic of Tapioca, Citrus Canker, Blast disease of Paddy.
2. Students should be trained to prepare the fungicide Bordeaux mixture & Tobacco decoction.

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SEMESTER-III

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EXPECTED OUTCOME

- ✎ The student can prepare micro preparations and identify the thallus and reproductive structures of lower plant groups like algae, fungi and lichen
- ✎ An awareness created among students about various microbes, structure and economic importance
- ✎ Students can use effectively the methodology to isolate and identify bacteria present in curd and root nodules
- ✎ Can identify various plant diseases, etiology of pathogens and control measures
- ✎ Able to prepare fungisides like tobacco decoction and Bordeaux mixture

SEMESTER-IV

BRYOLOGY, PTERIDOLOGY, GYMNOSPERMS AND PALAEOBOTANY

Course code : BO 1441, Number of credits : 3

Number of Contact Hours: Lecture- 54 Hours; Practicals-36 Hours Total- 90 Hrs

Distribution of Hours	Theory	Practical
Bryology	18 hrs	09 hrs
Pteridology	18 hrs	14 hrs
Gymnosperms	14 hrs	12 hrs
Paleobotany	04 hrs	01 hr
Total	54 hrs	36 hrs

Aim of the course: To create awareness about non flowering plant groups like Bryophytes, Pteridophytes and Gymnosperms.

Objectives:

- ☒ To familiarize the students characteristic features and evolutionary significance of Bryophytes, Pteridophytes and Gymnosperms.
- ☒ To generate awareness about lifecycle of Bryophytes, Pteridophytes and Gymnosperms.
- ☒ To impart knowledge about fossil formation and its significance

MODULE-1

Bryology 18hrs

1. Introduction and general characters, classification- Proskauer (1957)
2. Study of habit, thallus organization, vegetative and sexual reproduction and alternation of generation of the following types (Developmental details are not required)
Riccia, Marchantia, Anthoceros, Funaria
3. Economic importance of Bryophytes

Practical 09Hrs

1. *Riccia*- Habit- Internal structure of thallus- V.S.of thallus through archegonia, antheridia and sporophyte
2. *Marchantia*- Habit- thallus with Archegonial receptacle, Male receptacle V.S, Female receptacle V.S. T.S of thallus through gemma, Sporophyte V.S
3. *Anthoceros* – Habit with sporophyte, Sporophyte V S
4. *Funaria*- Habit V.S. of archegonial cluster, V.S of Antheridial cluster, Sporophyte V.S

MODULE -11

Pteridology 18hrs

1. Introduction: General characters morphological and phylogenetic classification

SEMESTER-IV

(Smith, 1955; Bold, 1957 and Zimmerman, 1959).

2. Study of the habitat habit, internal structure, reproduction and life cycle of the following types (Developmental details not required). *Psilotum*, *Selaginella*, *Pteris* and *Marselia*
3. General topics- Stellar evolution in Pteridophytes - Economic importance of Pteridophytes

Practical

14hrs

1. *Psilotum* -External features, Stem T.S., Synangium T.S
2. *Selaginella* - Habit , stem and rhizophore, T.S, V.S of strobilus, Megasporophyll and Microsporophylls
3. *Pteris* - Habit , Rachis T.S Sporophyll T.S, Prothallus
4. *Marselia* - Habit, Rhizome and Petiole T.S, Sporocarp T.S ,V.S

MODULE-III

Gymnosperms

14hrs

1. Introduction –General characters and classification of Gymnosperms (Sporne, 1965)
2. Study of the habit, anatomy, reproduction and life cycle of the following types (Developmental details are not required) *Cycas*, *Pinus* and *Gnetum*
3. Evolutionary trends in gymnosperms, Economic importance of Gymnosperms

Practical

12hrs

1. *Cycas*- T.S of leaf, T.S. of coralloid root. Micro and megasporophyll, V S of ovule
2. *Pinus*- T.S. of stem, T.S. of needle, male and female cone, V.S.
3. *Gnetum*-Habit, stem T.S (young and mature), leaf T.S, male and female strobilus, V.S of male and female cone, ovule V.S and seed.

MODULE –IV

Palaeobotany

04hrs

1. Geological time scale, Fossil formation, types of fossils.
2. Fossil Pteridophytes- *Rhynia*, *Lepidodendron*, *Lepidocarpon*. Fossil gymnosperms- *Lyginopteris*.

Practical

01hr

1. Fossil Pteridophytes- *Rhynia* stem, *Lepidodendron*, *Lepidocarpon*.
2. Gymnosperm-*Lyginopteris*

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SEMESTER-IV

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EXPECTED OUTCOME

- ✎ Students are able to make micropreparations of thallus and reproductive structures of as well as better understanding of the life cycle of selected members of Bryophytes, Pteridophytes and Gymnosperms
- ✎ Can understand the economic and ecologic importance of lower groups of plant kingdom
- ✎ Better understanding of fossilization and importance of Palaeobotany
- ✎ Identify various parts of fossil plants through micro slides

SEMESTER-V

ANGIOSPERM MORPHOLOGY, SYSTEMATIC BOTANY, ECONOMIC BOTANY, ETHNO BOTANY AND PHARMACOGNOSY

Course Code :BO1541, Number of Credits :4

Number of contact hours : Lecture: 72 hrs; Practical: 54 hrs; Total: 126 hrs

Distribution of Hours	Theory	Practical
Morphology	14 hr	9 hr
Systematic Botany	40 hr	36 hr
Economic Botany	12 hr	06 hr
Ethnobotany & Pharmacognosy	06 hr	03 hr
Total	72 hr	54 hr

Aim of the course: To understand classification, identification and preservation of angiosperms along with ethnobotanical importance.

Objectives:

- ✎ To introduce importance of morphological characters in classification and plant identification.
- ✎ To develop skill for herbarium preparation.
- ✎ To acquire knowledge about economic, ethnobotanical significance and pharmacognosy of plants

MODULE-I

Morphology

14hrs

1. Morphological description of a flowering plant: Plant habit, Stem (brief account), Leaf – morphotypes, phyllotaxy, Venation.
2. Various types of inflorescence including special types (Cyathium, Verticillaster, Hypanthodium, Coenanthium and Thyrsus) with examples;
3. Floral morphology- Flower-as a modified shoot, Flower parts, their arrangements, relative position, numeric- plan, cohesion, adhesion, symmetry of flower, aestivation types, placentation types. Description of a plant in technical terms, Floral diagram and floral formula
4. Fruit types: simple, aggregate and multiple. Seeds: albuminous and exalbuminous.

MODULE-II

Systematic Botany

05hrs

Definition, scope and significance of Taxonomy, Historical development of the systems of classification:

1. Artificial- sexual system of Linnaeus

SEMESTER-V

2. Natural - Bentham and Hooker (detailed account)
3. Phylogenetic- Engler and Prantl (Brief account only)
4. APG system- Brief account only

MODULE-III

04hrs

1. Basic rules of Binomial Nomenclature and International Code of Botanical Nomenclature (ICBN).
2. Importance of herbarium, Herbarium techniques and Botanical gardens. Brief account on the Modern trends in taxonomy; Chemotaxonomy and Molecular taxonomy

MODULE-IV

31 hr

A study of the following families with emphasis on the morphological peculiarities and economic importance of its members (based on Bentham & Hooker's system)

- | | | |
|------------------|--------------------|-------------------|
| 1. Annonaceae | 11. Rubiaceae | 21. Euphorbiaceae |
| 2. Nymphaeaceae | 12. Asteraceae | 22. Orchidaceae |
| 3. Malvaceae | 13. Sapotaceae | 23. Scitaminae |
| 4. Rutaceae | 14. Apocynaceae | 24. Liliaceae |
| 5. Anacardiaceae | 15. Asclepiadaceae | 25. Arecaceae |
| 6. Leguminosae | 16. Solanaceae | 26. Poaceae |
| 7. Combretaceae | 17. Acanthaceae | |
| 8. Myrtaceae | 18. Verbenaceae | |
| 9. Cucurbitaceae | 19. Lamiaceae | |
| 10. Apiaceae | 20. Nyctaginaceae | |

Practical

45hrs

1. Study on various types of inflorescences, Flowers and fruits with vivid record of practical work.
2. Students must be able to identify the angiosperm members included in the syllabus up to the level of families. Draw labeled diagram of the habit, floral parts, L S of flower, T S of ovary, floral diagram, floral formula and describe the salient features of the member in technical terms. (Minimum two plants from each dicot family and one from monocot family).
3. Students must submit practical records, Herbarium sheets (25 Nos: representing one sheet from each family) and Field book at the time of practical examination.
4. Field trips are to be conducted for three days either as continuous or one daytrips.

MODULE-V

a. Economic Botany

12hrs

1. Study of the major crops in Kerala with special reference to their Botanical description, morphology of the useful part and economic importance—
 - i. Coconut & Paddy
 - ii. Cereals— Wheat & Maize
 - iii. Millets- Ragi & Fox tail millet

SEMESTER-V

- iv. Pulses – Black gram, Green gram, Bengal gram
- v. Sugar yielding plants – Sugar Cane
- vi. Spices- pepper, cloves, cardamom
- vii. Beverages – Coffee, Tea
- viii. Fibre yielding plants – Cotton
- ix. Dye Yielding plants - Henna and *Bixa Orellana*
- x. Resins- Asafoetida
- xi. Tuber crops – Tapioca, Potato
- xii. Oil yielding plants- Sesame, ground nut
- xiii. Latex yielding plants- Rubber
- xiv. Medicinal plants - *Sida*, *Zingiber officinalis*, *Aloe vera* and *Vinca rosea*
- xv. Insecticide- Neem

Practical **06hrs**

Identify the economic products obtained from the plants mentioned under Economic Botany

b. Ethnobotany **03hrs**

1. Definition — importance, scope, categories and significance.
2. Study of various methods to collect Ethno botanical data.
3. Plant parts used by tribes in their daily life as food, clothing, shelter, agriculture and medicine.
4. Study of common plants used by tribes. *Aegle marmelos*, *Ficus religiosa*, *Cynadondactylon*, *Ocimum sanctum* and *Trichopus zeylanicus*

c. Pharmacognosy **03hrs**

1. Definition and scope of Pharmacognosy
2. Sources of crude drugs – roots, rhizome, bulb, corm, leaves, stems, flowers, fruits and seeds

Practical **03hrs**

1. Visit to a tribal area and collection of information on their traditional method of treatment using crude drugs
2. Observe the plants of ethno botanical importance

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SEMESTER-V

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EXPECTED OUTCOME

- ✗ Ability to identify different types of inflorescences, flowers and fruits, their arrangement and relative position.
- ✗ Familiarization of basic rules of Angiosperm classification and different types of classification.
- ✗ Preparation and maintenance of Herbarium.
- ✗ Identification of plants to their respective families.
- ✗ Understanding of ethnobotanical and pharmacological significance of plants.

SEMESTER-V

ENVIRONMENTAL STUDIES AND PHYTOGEOGRAPHY

Course code: BO 1542 Number of Credits : 4

Number of contact hours : Lecture: 90 hrs; Practical: 36 hrs; Total: 126 hrs

Distribution of Hours	Theory	Practical
Environmental Studies	72 hrs	36 hrs
Disaster Management	09 hrs	00 hrs
Phytogeography	09 hrs	00 hrs
Total	90 hrs	36 hrs

Aim of the course: To inculcate environmental awareness among students for protecting the Nature.

Objectives:

1. To create awareness about ecosystem and Natural resources.
2. To generate knowledge about importance of Biodiversity conservation
3. To understand the need to mitigate pollution and strategies for disaster management
4. To impart knowledge about phytogeographical regions

MODULE-I

Natural resources and its conservation

12hrs

5. Natural Resources - Renewable and Non-renewable - Land & Soil, Water, Energy, Minerals, Food and agriculture, Forests, Plants & Wild life resources. Degradation of natural resources - Land degradation, degradation of water resources, Loss of flora and fauna; Causes – population explosion, over exploitation, deforestation, agriculture mismanagement, desertification, overgrazing, soil erosion, mining, urbanization and industrialization- change in land use, depletion of water resources.
6. Conservation of Natural resources and sustainable life styles.
Land and soil- Afforestation, regeneration of wasteland
Water - Rain water harvesting, ground water dams
Energy - Promoting use of renewable resources-solar, tidal and wind; biodiesel, biofuels.
Forests- Reforestation, Community forestry programmes

MODULE- II

Ecosystems

22 hr

1. Ecosystems - Concept, definition, structure and function; components- biotic and abiotic; energy flow
2. Food chains -Food web & ecological Pyramids, biogeochemical cycles - Carbon and Phosphorous cycle
3. Ecological succession-Definition, primary and secondary succession, climax

SEMESTER-V

concept, hydrosere and xerosere.

4. Plant adaptations- Morphological, anatomical & physiological adaptations of – Hydrophytes, Xerophytes, Halophytes, Epiphytes, Parasites
5. Introduction- types, characteristic features, structure and functions of the following ecosystems.
6. Forest ecosystem, 2. Grassland ecosystem, 3. Desert ecosystem, 4. Aquatic ecosystems- Ponds, Streams, Rivers, Oceans, Estuaries (brief account only)

MODULE-III

Biodiversity and its conservation

18hrs

1. Introduction, Definition- genetic, species and ecosystem diversity; biodiversity index (Shannon index)
2. Hot-spots of biodiversity; India as mega-diversity nation.
3. Threats to biodiversity: land use changes & habitat destruction, poaching of wild life- hunting & export, Overuse of pesticides, invasive species.
4. IUCN, Red data Book; Extinct and Threatened species- endangered & Rare; Endemic species of Western Ghats.
5. Conservation of biodiversity: In-situ (National parks and Wild life sanctuaries) and Ex-situ conservation (botanical gardens); Biosphere Reserves & World Heritage Sites in India-Ramsar sites, Chilka lake.
6. Global initiatives in biodiversity conservation- Stockholm Conference, Montreal Protocol, Convention on Biological diversity Regional initiatives- Chipko movement, National Biodiversity Authority (NBA), Access and Benefit sharing (ABS), A brief account on conservation efforts in Kerala- Kerala State Biodiversity Board (KSBB), People Biodiversity Register.

MODULE-IV

Environmental pollution

20hrs

1. Definition, causes, effects and control measures of – 1. Air pollution, 2. Water pollution, 3. Soil pollution, 4. Marine pollution, 5. Noise pollution, 6. Thermal pollution.
2. Solid Waste Management- waste minimization, Recycling and Reuse, Consuming environment friendly products. E-waste management.
3. Environmental Legislations - Environment protection Act (1986); Air [prevention and control of pollution] Act (1981; Amended 1987); Water [prevention and control of pollution] Act (1974; Amended 1988); Wildlife Protection Act (1972); Forest conservation Act (1980). (Scope and relevance only)
4. Environmental Organisations –UNEP, IPCC, WWF, Central Pollution Control Board

MODULE-V

Disaster management

09hrs

1. Introduction, Definition and terminologies; scope and concept of disaster management.
2. Natural and Environmental disasters-a brief description of the following disasters-

SEMESTER-V

earth quake, flood, coastal disasters, landslides, tsunami(role of mangroves in controlling tsunami disaster), cyclone, dam collapse, nuclear disaster, chemical disaster, biological disaster.

3. Environmental Issues - Global warming and sea level rise, Acid rain, Ozone layer depletion- causes and effects.
4. Disaster management – four phases – mitigation, preparedness, responses, recovery. Emergency procedures and warning systems, application of GIS (brief account only).

MODULE-VI

Phytogeography

9 hrs

1. Concept & definition, species distribution- continental drift, continuous and discontinuous distribution.
2. Vegetation in India – Forests- tropical, temperate, sholas, sub alpine, alpine, mangroves & Grass lands.
3. Phytogeographical regions of India - Western and eastern Himalayas, Desert, Western Ghats, Deccan Peninsula, Gangetic Plain, North East India, Coasts & Islands

Practical

36hrs

1. Visit a local polluted site and report major pollutants.
2. Visit a mangrove vegetation and report diversity
3. Study of ecological and anatomical modifications of Xerophytes, Hydrophytes,
4. Halophytes, Epiphytes and Parasites.
5. Observation and study of different ecosystems mentioned in the syllabus.
6. Phytogeographical regions of India- Photos/Diagrams

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EXPECTED OUTCOME

- ☒ Develops awareness about natural resources, its conservation and importance of sustainable lifestyles.

SEMESTER-V

- ✎ Understands and identify different ecosystems and ecosystem processes.
- ✎ Develops deep understanding about biodiversity and importance of its conservation
- ✎ Develops skills to identify polluted sites, its major pollutants and recognize the need to mitigate environmental pollution
- ✎ Awareness about different types of disasters and to adopt strategies to overcome and reduce the impact
- ✎ Identify the importance of phytogeographical sites in India

SEMESTER-V

CELL BIOLOGY, GENETICS AND EVOLUTIONARY BIOLOGY

Course Code : BO 1543, Number of credits : 3

Number of contact hours : Lecture: 72 hrs; Practical: 36 hrs; Total: 108 hrs

Distribution of Hours	Theory	Practical
Cell biology	25 hrs	12 hrs
Genetics	36 hrs	24 hrs
Evolutionary biology	11 hrs	00 hrs
Total	72 hrs	36 hrs

Aim of the course: To familiarize cellular organelles, Classical Genetics and importance of Evolution.

Objectives:

- ✎ To create awareness about cellular organelles.
- ✎ To develop skills to identify cell stages and workout problems in classical genetics.
- ✎ To introduce different theories of evolution

MODULE-I

Cell Biology

25hrs

1. History and progress of cell biology
2. Ultra structure and functions of the cell components and organelles Cell wall; The cell membrane, Endoplasmic reticulum, Ribosomes, Golgi apparatus, Lysosomes, Peroxisomes, Vacuole, Mitochondria, Chloroplast & Nucleus.
3. The chromosomes- Chromosome morphology- Eukaryotic chromosomes and its organization. Chromatin - composition and structure; hetero chromatin and euchromatin; Chemical organization. Nucleoproteins – histones and non – histones. Nucleosome model of DNA organization.
4. Special types of chromosomes- Salivary gland, Lamp brush and B chromosomes
5. Variation in Chromosome number (Numerical aberrations)- aneuploidy and Euploidy- haploidy, polyploidy- significance
6. Variation in Chromosome structure (Structural aberrations) - deletion, duplication, inversion and translocation; significance
7. Mitosis and Meiosis. cell cycle : Significance of mitosis and meiosis

Practical

12hrs

1. Make acetocarmine squash preparation of onion root tip and to identify different stages of mitosis.
2. Make squash preparation of the flower buds of any of the following plants. *Rhoeo, Capsicum* (To identify Meiosis) Demonstration only

SEMESTER-V

MODULE-II

Classical Genetics

36hrs

1. Mendelian Genetics- Mendel and his experiments, Mendel's success, Mendelian principles, Mendelian ratios, monohybrid and dihybrid crosses, back cross and testcross
2. Genetics after Mendel- Modified Mendelian ratios; Incomplete dominance – Flower color in *Mirabilis* ; Interaction of genes- Comb pattern in poultry. 9:3:3:1. Epistasis - Recessive. Coat color in mice. 9:3:4; Dominant epistasis. Fruit colour in summer squash. 12:3:1; Complementary genes. Flower color in *Lathyrus* 9:7; Duplicate gene with cumulative effect. Fruit shape in summer squash. 9:6:1; Duplicate dominant genes in shepherd's purse. 15:1; Inhibitory factor. Leaf color in Paddy. 13:3
3. Multiple alleles-General account. ABO blood group in man. Rhfactor.
4. Quantitative characters- General characters of quantitative inheritance, polygenic inheritance; Skin color in man, ear size in Maize.
5. Linkage and crossing over- Linkage and its importance, linkage and independent assortment. Complete and incomplete linkage. Crossing over – a general account, two point, three point cross. Determination of gene sequence. Interference and coincidence. Mapping of chromosomes.
6. Sex determination- Sex chromosomes, chromosomal basis of sex determination XX- XY, XX-XO mechanism. Sex determination in higher plants (*Melandrium album*) Sex chromosomal abnormalities in man. Klinefelter's syndrome, Turner's syndrome. Sex linked inheritance. Eye colour in *Drosophila*, Hemophilia in man.
7. Extra nuclear inheritance General account, maternal influence. Plastid inheritance in *Mirabilis*. Shell coiling in snails, kappa particle in *Paramecium*.

Practical

24hrs

Work out problems in

1. Monohybrid cross (Dominance and incomplete dominance)
2. Dihybrid cross (Dominance and incomplete dominance)
3. Gene interactions (All types of gene interactions mentioned in the syllabus)
 - a. Recessive epistasis 9: 3: 4.
 - b. Dominant epistasis 12: 3:1
 - c. Complementary genes 9:7
 - d. Duplicate genes with cumulative effect 9: 6:1
 - e. Inhibitory genes 13:3
 - f. Duplicate dominant gene 15: 1
 - g. Comb pattern in poultry 9:3:3:1
4. Linkage and crossing over
5. Two point and three point crosses
6. Construction of genetic map.

MODULE-III

Evolutionary Biology

11hrs

1. Progressive and Retrogressive evolution.
2. Parallel and Convergent evolution.

SEMESTER-V

3. Micro and Macro evolution.
4. Theory of Lamarck, Wiesman and De Vries, Darwinism, Neo-Darwinism
5. Isolation, Mutation, Genetic drift, Speciation
6. Variation and Evolution – Hybridization and Evolution – Polyploidy and evolution– Mutation and Evolution.

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SEMESTER-V

EXPECTED OUTCOME

- ✎ Students have a better understanding of cell structure and cell organelles
- ✎ Can prepare microslides of cell divisions and identify various stages of mitosis and meiosis
- ✎ Able to workout problems in classical genetics, modified mendelian ratios and population genetics
- ✎ Able to understand genetic diseases and their inheritance
- ✎ Understand evolutionary principles, theories and methods of speciation

SEMESTER-VI

PLANT PHYSIOLOGY AND BIOCHEMISTRY

Course code : BO 1641 Number of Credits : 4

Number of contact hours: Lecture: 90 hrs; Practical: 36 hrs; Total: 126 hrs

Distribution of Hours	Theory	Practical
Plant Physiology	60 hrs	20 hrs
Biochemistry	30 hrs	16 hrs
Total	90 hrs	36 hrs

Aim of the course: To create awareness about physiological and biochemical aspects of growth & metabolism

Objectives:

- ✎ To understand physiology of absorption, photosynthesis and respiration.
- ✎ To study physiological responses in growth, movements and flowering of plants
- ✎ To generate awareness about biomolecules.
- ✎ To develop skill for testing of biomolecules

MODULE-I

20hrs

1. General introduction: physiological processes, their significance and applications.
2. Water relations of plants: Importance of water to plant life.
 - a. Absorption of water- organs of absorption, root and root hair. Physical aspects of absorption- imbibition, diffusion and osmosis. Plant cell as an osmotic system; water potential and osmotic potential. Plasmolysis and its significance, practical applications. Mechanism of water absorption – active and passive absorption, root pressure. Pathway of water across root cells.
 - b. Ascent of sap- vital and physical theories.
 - c. Loss of water from plants : transpiration-cuticular, lenticular and stomatal mechanism
Theories – starch sugar hypothesis, potassium - ion theory. Significance of transpiration - guttation, anti-transpirants, factors affecting transpiration.
3. Mineral nutrition: Gross chemical analysis of the plant body, ash analysis, criteria for essentiality of elements, macro and micro elements, role of essential elements and their deficiency symptoms. Culture methods - sand culture, hydroponics and aeroponics. Mechanism of mineral absorption (a) passive absorption- ion exchange and Donnan equilibrium (b) active absorption- carrier concept, Lundegardh hypothesis.

MODULE-II

20hrs

1. Photosynthesis: Introduction, significance and general equation; Photosynthetic apparatus, structure and function of chloroplast, quantasomes - solar spectrum and its importance - Fluorescence and phosphorescence; Red drop, Emerson effect; Two pigment systems; raw material for photosynthesis; Mechanism of

SEMESTER-VI

photosynthesis- Light reaction - cyclic and non cyclic photophosphorylation; Hill reaction - Dark reaction: Calvin cycle; Comparative study of C₃, C₄ and CAM plants; Photorespiration Bacterial photosynthesis and chemosynthesis (Brief account only)

2. Factors affecting photosynthesis - Law of limiting factor.
3. Respiration: Introduction, definition and significance and general equation. Respiratory substrate, types of respiration- aerobic and anaerobic. Aerobic respiration - glycolysis, Krebs's cycle, terminal oxidation. Anaerobic respiration – fermentation: alcoholic and lactic acid fermentation. Energy relation of respiration - R .Q and its significance - Factors affecting respiration.

MODULE-III

20hrs

1. Translocation of solutes: Path way of movement, phloem transport, mechanism of transport - Munch hypothesis, protoplasmic streaming theory - activated diffusion hypothesis, electro osmotic theory.
2. Nitrogen metabolism: Source of nitrogen - Biological nitrogen fixation – symbiotic and asymbiotic. Nitrogen fixation by blue green algae - rotation of crops. Nif genes - Leghaemoglobin.
3. Growth: Phases of growth - vegetative and reproductive growth - growth curve - plant growth regulators - Auxins, Gibberellins, Cytokinins, Ethylene, Abscissic acid - synthetic plant hormones - practical applications. Senescence and abscission, Photoperiodism
4. Vernalization - phytochrome and its significance. Physiology of bud and seed dormancy, germination.
5. Plant movements: Tropic and nastic movements. Circadian rhythm and biological clock.
6. Stress physiology: water stress, salt stress.

Practical

20hrs

1. Water potential of onion peel / *Rhoeo* peel by plasmolytic method.
2. Imbibition of water by different types of seeds.
3. Effect of temperature on permeability.
4. Papaya petiole osmoscope.
5. Determination of stomatal index.
6. Determination of water absorption and transpiration ratio.
7. Measurement of rate of transpiration using Ganong's potometer or Farmer's potometer.
8. Evolution of oxygen during photosynthesis.
9. Light screen experiment
10. Measurement of photosynthesis by Wilmott's bubbler.
11. Evolution of CO₂ during respiration.
12. Ganong's respirometer and measurement of R.Q
13. Alcoholic fermentation using Kuhn's fermentation vessel
14. Geotropism using clinostat
15. Measurement of growth using Arc auxanometer.

SEMESTER-VI

MODULE-IV

Biochemistry

15hrs

1. Molecules and life.
2. Carbohydrates - Classification, occurrence, structure and functions of monosaccharides (glucose and fructose), oligosaccharides (sucrose and maltose), polysaccharides (starch and cellulose), synthesis of glycosidic bonds – Enzymatic hydrolysis of glycosidic bonds – amylases and invertases.
3. Amino acids- classification based on polarity, structure - Amphoteric property of Amino acids
4. Peptide formation – Amino acid metabolism – reductive amination and transamination
5. Proteins – Structure, classification, properties and function; Role of bonds in stabilizing protein structure - hydrolysis of proteins.

MODULE-V

15hrs

1. Lipids- classification – Simple lipids- fats & oils, waxes; Compound lipids- phospholipids, sphingo lipids and glycolipids; Derived lipids- Cholesterol and terpenes; Fatty acids – Alpha- oxidation and Beta-oxidation; Synthesis of ester bonds.
2. Enzymes - general account - structure, classification and nomenclature (recommended by Commission on Enzymes); Mechanism of enzyme action - inhibition of enzymes - regulation of enzymes - allosteric inhibition - Isoenzymes, coenzymes and cofactors
3. Secondary Plant Products – Introduction – classification and function [General account]
4. Phytochemicals- Alkaloids, terpenoids, phenolics, flavonoids

Practical

16hrs

1. Qualitative test for carbohydrates - Molisch's test, Benedict's test (for reducing sugar)
2. Iodine test for starch
3. Test for proteins – Biuret test

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SEMESTER-VI

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Expected outcome of the Course

- ✎ Students get a clear understanding of the basic concepts of Physiology and Biochemistry.
- ✎ Understands photosynthesis, respiration, plant growth regulators, nitrogen metabolism and stress physiology
- ✎ Familiarization of basic physiological practical procedures.
- ✎ Students get the basic knowledge about the macromolecules and their overall role in cell metabolism; and secondary plant products.
- ✎ Identification of protein, reducing and non reducing sugar by qualitative tests.

SEMESTER-VI

MOLECULAR BIOLOGY, GENERAL INFORMATICS & BIOINFORMATICS

Course code : BO 1642, Number of credits : 4

Number of contact hours: Lecture: 72 hrs; Practical: 36 hrs; Total: 108 hrs

Distribution of Hours	Theory	Practical
Molecular Biology	36 hrs	9 hrs
General informatics	18 hrs	9 hrs
Bioinformatics	18 hrs	18 hrs
Total	72 hrs	36 hrs

Aim of the course: To develop knowledge about molecular biology of genetic material and gene expression along with information technology and biological databases

Objectives:

- ✗ To generate awareness of genetic material and gene expression.
- ✗ To get an overview of information technology
- ✗ To develop skill for using internet, biological databases and molecular visualization tools

MODULE-I

Molecular Biology

36hrs

1. DNA as genetic material- experimental evidence- Griffith's experiment on Bacterial transformation, Avery's experiment, Hershey-Chase Experiment.
2. DNA- Chemical Composition, Chargaff's rules, molecular structure of DNA- Watson & Crick's Double Helical Model of DNA, Salient features of double helix, Biological Significance of Double Helical Model of DNA; Forms of DNA - A, B and Z forms; Satellite and repetitive DNA
3. Replication of DNA in prokaryotes- An overview, General principles and features, Semi conservative model- Meselson and Stahl experiment; Leading strand and lagging strand synthesis, Okazaki fragments, replication fork and origin of replication; Unidirectional and Bidirectional replication; Replisome. Enzymology of replication: topoisomerase, helicase, primase, polymerase and ligase; DNA repairing mechanism-photoreactivation. Replication of DNA in eukaryotes (brief account only).
4. RNA structure- Structure, Properties and functions of tRNA, mRNA and rRNA; Genetic code.
5. Synthesis of protein: Transcription; RNA modifications- introns, exons, removal of introns, spliceosome; Translation -Central dogma-reverse transcription
6. Regulation of gene expression in prokaryotes and eukaryotes-lac operon;

SEMESTER-VI

transcriptional gene regulation in eukaryotes-promoters, enhancers, transcription factors; RNA interference.

7. Concept of gene-Units of a gene, cistron, recon, muton; Types of genes- House keeping genes (constitutive genes), Luxury genes (non constitutive genes), overlapping genes.
8. Transposable genetic elements- General account, Characteristic, Transposons (jumping genes), Cellular oncogenes (general account only).

Practical

9hrs

1. Study of semi-conservative replication of DNA through micrographs/ schematic representations.
2. Practice problems in molecular biology based on DNA structure and replication

MODULE-II

General Informatics

18hrs

1. Overview of Information Technology: Features of the modern personal Computer and peripherals, computer network and internet, overview of operating systems- Windows & Linux, and major application software's - Excel, Power point, MSword.
2. Knowledge skill for Higher Education: Data information and knowledge, knowledge management- Internet as a knowledge repository, academic search techniques, creating your cyber presence, open access initiatives, open access publishing models, basic concepts of IPR, copy rights and patents, plagiarism, introduction to use of IT in teaching and learning, case study of educational software, Academic services- INFLIBNET, NICNET and BRNET.
3. Social Informatics: IT and Society- issues and concerns- digital divide, IT and development, new opportunities and new threats, Cyber ethics, Cyber crime, Security, privacy issues, cyber addictions, Information overload, Health issues, guidelines for proper usage of computers, internet and mobile phones. Localization issues-IT and Regional languages-IT for the disabled, the free software debate.

Practical

9hrs

1. Create, Copy and Save a document with Header, Footer, Page Number, Date and Time using Word processing software.
2. Insert a table in the above Document
3. Prepare the mark list of students in a class using Excel
4. Prepare five slides each using power point with different design templates

MODULE-III

Bioinformatics

18hrs

1. Introduction: Definition, Origin of concept of Bioinformatics; Brief history, Importance of bioinformatics, Wet lab and Weblab.
2. Basics of Genomics, Proteomics & Comparative genomics
3. Biological databases:

SEMESTER-VI

- ✎ Nucleic acid databases (Eg: EMBL, Gen Bank, DDBJ)
 - ✎ Protein sequence databases. Eg: PIR, SWISS PROT, UNIPROT
 - ✎ Brief account on Model/organism databases, Biodiversity data bases
 - ✎ Protein structure databank- PDB
4. Gene sequence, Sequence analysis and alignment (brief account only), Pair wise sequence alignment, multiple sequence alignment, Sequence Alignment Tools: BLAST, CLUSTAL X
 5. Bioinformatics in relation to Biomolecular structure.
 6. Molecular visualization- use of Rasmol
 7. Molecular Phylogeny and Phylogenetic trees- Advantages of Molecular phylogeny and phylogenetic analysis- PHYLIP

Practical

18 hrs

1. Molecular visualization using Rasmol
2. Blast Search
3. Students should access Gene databases, download and take a print out of any one of gene sequences
4. Students are expected to work with at least any one of the commercial / scientific packages, to explore the WEB and able to find, recognize, download, install and use software in various areas useful to the research in Biology.

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SEMESTER-VI

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 18. Selzer PM, Marhofer RJ, Rohwer A (2009) Applied Bioinformatics. Springer-Verlag Berlin Heidelberg, Germany
 19. Teresa Attwood and David Parry-Smith (1999) Introduction to Bioinformatics Prentice Hall
 20. Twymann, R.M. (1998) Advanced molecular biology, Viva books NewDelhi.

WEB RESOURCES

- ✎ www.fgcu.edu/support/office2000
- ✎ www.openoffice.org *Open office officialwebsite*
- ✎ www.microsoft.com/office *MS Officewebsite*
- ✎ www.lgta.org *Office onlinelessons*
- ✎ www.learntheneth.com *WebPrimer*
- ✎ www.computer.org/history/timeline
- ✎ www.computerhistory.org
- ✎ <http://computer.howstuffworks.com>
- ✎ <http://vmoc.museophile.org> *Computerhistory*
- ✎ www.dell.com *DellComputers*
- ✎ www.intel.com *Intel*
- ✎ www.ibm.com *IBM*
- ✎ www.keralaitmission.org *Kerala Govt. ITDept.*
- ✎ www.technopark.org
- ✎ www.studentworkzone.com/question.php?ID=139

Expected outcome

- ✎ Understands DNA as genetic material, develops awareness about chemical composition and different types of DNA including their replication method.
- ✎ Students understand various molecular aspects of gene expression and regulation of genes
- ✎ Develops awareness about various academic services applied for their studies
- ✎ Awareness about features of a computer, different application and system software.
- ✎ Recognizes the need for safe use of internet and also become aware about health issues related to over usage of computers and mobile phones as well as cyber crimes and cyber laws.
- ✎ Students will be familiarized to molecular phylogeny, Biological Databases, Sequence analysis, Genomics, Proteomics & Comparative genomics

SEMESTER-VI

HORTICULTURE, PLANT BREEDING & RESEARCH METHODOLOGY

Course code : BO 1643, Number of credits : 4

Number of contact hours: Lecture: 72 hrs; Practical: 36 hrs; Total: 108 hrs

Distribution of Hours	Theory	Practical
Horticulture	35 hrs	20 hrs
Plant breeding	20 hrs	16 hrs
Research methodology	17 hrs	00 hrs
Total	72 hrs	36 hrs

Aim of the course: To introduce horticultural techniques and plant breeding along with awareness in research methodology

Objectives:

- ✎ To get an awareness in principles and methods of gardening
- ✎ To understand plant breeding techniques and develop skill for hybridization.
- ✎ To get knowledge about research methodology and preparation of projects

MODULE- I

Horticulture

18hrs

1. Introduction - Divisions of horticulture- Importance and scope of horticulture.
2. Principles of garden making- types of pots and containers- Potting mixture and potting media – soil, sand, peat, sphagnum moss, vermiculite- Soil types, Soil preparation- Irrigation methods
3. Propagation methods- Cuttings, Layering – Air layering, Ground layering (Tip, Trench and Compound), Budding – T- budding, Grafting – Approach grafting, Bridge grafting, whip and tongue grafting.
4. Garden tools and implements- Lawn mower, hand trowel, nursery spade, spade fork, garden hoe, weeder, tillers
5. Manures and fertilizers- Farmyard manure, compost, vermi compost and biofertilizers; Chemical fertilizers – NPK; Time and application of manures and fertilizers- Foliar sprays
6. Irrigation Methods–Surface, Drip irrigation, Sprinkler irrigation

MODULE- II

17hrs

1. Components of Garden- Landscaping principles; Lawns, Trees, shrubs and shrubberies, climbers and creepers, Flower beds and borders, ornamental hedges, edges, Drives, roads, walks and paths, Carpet beds, topiary, trophy, rockery, Conservatory or greenhouses
2. Indoor garden, Roof garden (Brief account only)

SEMESTER-VI

3. Bonsai
4. Flower Arrangement- Containers and requirements for flower arrangements- Free style, Shallow and Mass arrangement- Japanese- Ikebana. Dry flower arrangement

Practical

20hrs

1. Familiarise the garden tools and implements mentioned in the syllabus
2. Students must be trained to do Cutting/ layering/ grafting/budding.
3. Visit to a Botanical garden under the guidance of the teacher is recommended

MODULE-III

20hrs

Plant breeding

1. Introduction, objectives in plant breeding- - Important national and international plant breeding Institutes
2. Plant introduction. Agencies of plant introduction in India, Procedure of introduction - Acclimatization -Achievements.
3. Selection - mass selection, pure line selection and clonal selection. Genetic basis of selection and methods.
4. Hybridization: Procedure of hybridisation, inter generic, inter specific, inter varietal hybridisation with examples. Composite and synthetic varieties.
5. Heterosis and inbreeding depression- genetic basis; male sterility
6. Mutation breeding – method – achievements in India.
7. Polyploidy breeding
8. Breeding techniques and achievements with reference to the following crops in India:
(a) Rice (b) Wheat (c) Potato (d) Coconut

Practical

16hrs

1. Techniques of emasculation and hybridization of any bisexual flower.

MODULE-IV

17hrs

Research Methodology

1. Introduction; Need for research; Stages of Research – Definition of problem, execution of the problem, interpretation of results
2. Characteristics of Research, Types of research- Qualitative & quantitative.
3. Experimental design, components of experimental designs- Randomized blocks, completely randomized designs.
4. Preparation of a project report : Data analysis and consolidation of photographs, illustrations, tables and graphs, Title, introduction, review of literature, materials and methods, results, discussions, summary, references, acknowledgements; Bibliography – Method of citing and arrangement of references.

REFERENCES

1. Acquaah G (2007). Principles of Plant Genetics and Breeding, Blackwell Publishing Ltd. USA.
2. Allard RW (1999). Principles of Plant Breeding (2nd Edition), John Wiley and Sons.

SEMESTER-VI

3. Arora J.S (1990). Introductory Ornamental Horticulture, Kalyani Publications
4. Bailey L.H (1901). The Standard Cyclopaedia of Horticulture Volume 1,2 and 3, Macmillan Publications.
5. Bose T.K and Mukerjee D (1987). Gardening in India, Oxford Book House
6. Chauhan V.S (1972). Vegetable Production in India, Ram Prasad & Sons
7. Gupta S N (2010) Instant Horticulture, Jain Brothers publishing
8. Gurumani N. (2006) Research Methodology For Biological Sciences, MJP Publ.
9. Kothari C R & Garg C (2014) Research methodology methods and techniques, New Age international publishers
10. Kumar N (1989). Introduction to Horticulture, Rajalakshmi Publications
11. Manibhushan Rao K (2005). Text Book of Horticulture, Macmillan Publications
12. Shujnmoto, (1982). The Essentials of Bonsai, David & Charles ,Newton
13. Singh B D (2015). Plant Breeding, Kalyani publishers

Expected outcome

- ✗ Students able to identify and use various horticultural implements
- ✗ Can propagate plants through grafting, budding and layering & can prepare manures, fungicides etc
- ✗ Can effectively do plant breeding methods and understands their practical application in betterment of food crops
- ✗ Can devise an experimental design and carry out a project
- ✗ Students trained about various steps for the conduct of a research project and write a project report

OPEN COURSES

OPEN COURSE – I

Offered to the students of other disciplines choosing Botany open course

Semester	Course Code	Title of the Course	Contact hrs/week	Credits
V	BO1551.1	Horticulture	3	2
	BO1551.2	Mushroom Cultivation and Marketing		
	BO1551.3	Forestry		

OPEN COURSE –II

Offered to the students of Botany

Semester	Course Code	Title of the Course	Contact hrs/week	Credits
VI	BO 1651	Biotechnology & Nanobiotechnology	3	2

OPEN COURSE –I (A)

HORTICULTURE

Course code : BO1551.1, Number of credits : 2

Number of contact hours : Lecture: 54 hrs

Aim of the course: To develop knowledge about principles of gardening, propagation and flower arrangement

Objectives:

- ✎ To introduce horticultural methods
- ✎ To familiarize propagation methods in plants.
- ✎ To study types of gardens and flower arrangements

MODULE–I

10hrs

1. Introduction
2. Divisions of horticulture
3. Importance and scope of horticulture.
4. Principles of garden making
5. Types of pots and containers
6. Potting mixture and potting media – soil, sand, peat, sphagnum moss, vermiculite
7. Soil types, Soil preparation
8. Irrigation methods- drip irrigation and sprinkler irrigation
9. Hydroponics

MODULE–II

12hrs

I Propagation methods

1. Cuttings- root, stem, leaf
2. Layering – Air layering, Ground layering (Tip, Trench and Compound)
3. Budding – T-budding
4. Grafting – Approach grafting, Bridge grafting, whip and tongue grafting.
5. Garden tools and implements
6. Manures and fertilizers
7. Farmyard manure, compost, vermi compost and biofertilizers.
8. Chemical fertilizers –NPK.
9. Time and application of manures and fertilizers.
10. Foliar sprays

MODULE- III

12hrs

1. Components of Garden
2. Lawns and landscaping Trees, shrubs and shrubberies, climbers and creepers
3. Flower beds and borders, ornamental hedges, edges, Drives, roads, walks and paths , Carpet beds, topiary, trophy, rockery
4. Conservatory or green houses, Indoor garden, Roof garden
5. Bonsai

OPEN COURSE –I (A)

MODULE- IV

12hrs

1. Flower Arrangement
2. Containers and requirements for flower arrangements Free style, Shallow and Mass arrangement
3. Japanese –Ikebana
4. Bouquet and garland making
5. Dry flower arrangement
6. Harvesting Methods- Anthurium and Orchid &Storage
7. Marketing of Fruits, vegetables and flowers
8. Preservation and processing of fruits and vegetables

MODULE- V

8hrs

1. Growth regulators in horticulture
 2. Rooting hormones , Growth promoters , Flower induction, Parthenocarpy
 3. Common diseases of Mango and Tomato
 4. Plant protection- Weedicides, Fungicides, Pesticides
- Field Study:** Visit to a Botanical garden under the guidance of the teacher is encouraged.

REFERENCES

1. Arora J.S (1990). Introductory Ornamental Horticulture, Kalyani Publications
2. Bailey L.H (1901). The Standard Cyclopaedia of Horticulture Volume 1,2 and 3, Macmillan Publications.
3. Bose T.K and Mukerjee D (1987). Gardening in India, Oxford Book House
4. Chauhan V.S, (1972). Vegetable Production in India, Ram Prasad & Sons
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6. Kumar N (1989). Introduction to Horticulture, Rajalakshmi Publications
7. Manibhushan Rao K (1991). Text Book of Horticulture, Macmillan Publications
8. Shujnrrnoto, (1982). The Essentials of Bonsai, David & Charles, Newton
9. Rajesh Kumar, Kaushal Kumar Misra, (2014) Fundamentals of Horticulture, Biotech publishers
10. Singh J (2017) Fundamentals of Horticulture, Kalyani publishers

Expected outcome

- ☒ Students are familiarized in horticulture implements and methods of gardening
- ☒ Better understanding of commercial horticulture, flower arrangement, cut flowers
- ☒ Can understand about land scaping, fertilizers and Plant protection

OPEN COURSE -I (B)

MUSHROOM CULTIVATION AND MARKETING

Course code : BO1551.2, Number of credits : 2

Number of contact hours : Lecture: 54 hrs

Aim of the course: To create awareness about cultivation methods for mushroom and its economic aspects

Objectives:

- ✘ To get knowledge about nutritional and medicinal values of mushrooms
- ✘ To understand methods of mushroom cultivation.
- ✘ To give insight into the economic significance of mushroom cultivation

MODULE- I 12hrs

- ✘ History and introduction: Edible mushrooms and Poisonous mushrooms.
- ✘ Systematic position, morphology, distribution, structure of *Agaricus* and *Pleurotus*.

MODULE- II 08hrs

- ✘ Nutritional value, medicinal value and advantages- types- milky, straw, button and poisonous mushrooms

MODULE- III 12hrs

1. Cultivation: Paddy straw mushroom – substrate, spawn making. Methods – bed method, polythene bag method, field cultivation.
2. Oyster mushroom cultivation –Substrate, spawning, pre-treatment of substrate. Maintenance of mushroom.
3. Cultivation of white button mushroom – Spawn, composting, spawning, harvesting.

MODULE- IV 12hrs

1. Diseases- Common pests, disease prevention and control measures- *Agaricus*, *Pleurotus* and *Volverilla*.
2. Processing - Blanching, steeping, sun drying, canning, pickling, freeze drying.
3. Storage – short term and long term storage.

MODULE-V 10hrs

1. Common Indian mushrooms.
2. Production level, economic return, Foreign exchange from Mushroom cultivating countries and international trade.

Field Study: Visit to a mushroom cultivating Laboratory

REFERENCES

1. Harander Singh. (1991). Mushrooms- The Art of Cultivation- Sterling Publishers.
2. Indian Journal of Mushrooms. Published by I.M.G.A. Mushroom Research Laboratory. College Agriculture, Solan

OPEN COURSE -I (B)

3. Kaul T N (2001). Biology and conservation of mushrooms. Oxford and IBH publishing company N.Delhi
4. Pandey B P (1996). A textbook of fungi. Chand and Company NewDelhi.
5. Pavel Kalc (2016) Edible mushrooms, Chemical composition and nutritional value, Elseveir book aid international

Expected outcome

- ✎ Identify mushrooms, structure and mode of propagation
- ✎ Understand commercial mushroom cultivation, marketing and their nutritional value
- ✎ Better understanding of methods of processing and storage of mushrooms

OPEN COURSE -I (C)

FORESTRY

Course code : BO1551.3, Number of credits : 2

Number of contact hours : Lecture; 54 hrs

Aim of the course: To enlighten students about forest types and forest produce

Objectives:

- ✎ To get a knowledge about types of forest and silviculture
- ✎ To get knowledge about forest produce
- ✎ To understand agroforestry

MODULE - I

10hrs

1. General introduction to forests- Types of Forest- Natural and Manmade; Tropical, temperate, evergreen semi- evergreen, deciduous; Monoculture, multipurpose, social and industrial. Forest and gene conservation.

MODULE - II

16hrs

1. Silviculture- concept and scope of study of natural and artificial regeneration of forests. Clear felling, uniform shelter, wood selection, coppice and conservation systems. Silviculture of some of the economically important species in India such as *Azadirachta indica*, *Tectona grandis*, *Eucalyptus indica*, *Mahagoni mahagoni*, *Dalbergia sisso* and *Santalum album*, *Artocarpus heterophyllus*, *Hevea brasiliensis*.
2. Wood: Homogenous and heterogenous- spring and autumn wood- Porous and non porous wood- Heart and sap wood.
3. Relevance of wood anatomical studies in Kerala- Identification of wood- preparation of key and their uses.

MODULE - III

10hrs

1. Social and agro forestry. Selection of species and role of multipurpose trees. Food, fodder and energy.
2. Social forest- Avenue plantation. Sacred plants- definition, importance of sacred trees like *Ficus religiosa*, *Emblica officinalis*, *Aegle marmelos*.

MODULE - IV

09hrs

1. Seed orchards, seed dormancy- Types of dormancy, physical and chemical methods to overcome seed dormancy.
2. Forest laws- necessity, General principles, Indian forest act 1927 and their amendment.

MODULE - V

09hrs

1. Forest resources and utilization. Forest products- timber, pulp wood, secondary timbers, non timber forest products (NTFPs).
2. Definition and scope (brief outline) - Gums, resins, fibers, oil seeds, nuts, rubber, canes and bamboos, medicinal plants, charcoal. Lac collection and marketing.

OPEN COURSE -I (C)

Field Study

1. Identification of wood using key: Teak, Jack wood, Mahogany, Rubber, *Azadirachta*, Eucalyptus.
2. Visit to a plywood factory to have knowledge of wood based industry.

REFERENCES

1. A Hand book of Kerala Timbers- KFRI, Trichur.
2. Anil Kumar Dhiman. (2003). Sacred plants and their medicinal uses. Daya publishing house, NewDelhi
3. B.S. Chundawat and S.K.Gautham. (1996). Text book of Agro forestry. Oxford and IBH Publishing House, NewDelhi
4. Kollmann and Cote (1988). Wood science and Technology. Vol.I & II Springer verlag.
5. Parthiban K T (2016) Forestry, Competition Tutor publishers
6. Praveen Dhar T.(2018). Introduction to Forestry Narendra Publishing House, New Delhi
7. Sagreiya, K.P. (1994). Forests and Forestry (Revised by S.S. Negi). National book trust. NewDelhi.
8. Sharma P.D. (2004). Ecology and Environment. Rastogi publications, Meerut
9. Singh M.P. and Vinita Vishwakarma.(1997). Forest environment and Biodiversity. Daya publishing house, NewDelhi.
10. Tiwari K.M. (1983). Social forestry in India.
11. Tribhawan Mehta, (1981). A handbook of forest utilization. Periodical Expert Book Agency, NewDelhi.

Expected outcome

- ✎ Identification of forest types,Forest products and their utilisation and common forest trees of Kerala
- ✎ Understanding timber yielding plants and methods of identification of timber
- ✎ Understanding of germination of seeds of forest trees, methods to break dormancy of seeds, forest act and methods of preserving forests
- ✎ Get knowledge about silviculture and Social forestry

SEMESTER-VI

OPEN COURSE-II - ELECTIVE BIOTECHNOLOGY AND NANO BIOTECHNOLOGY

Course code : BO1651, Number of credits : 2

Number of contact hours : Lecture: 54 hrs

Aim of the course: To impart knowledge about Biotechnology and Nanobiotechnology

Objectives:

- ✎ To introduce plant biotechnology, tissue culture and rDNA technology
- ✎ To give insight into applications in industrial biotechnology and nano biotechnology

MODULE-I

12hrs

Biotechnology

1. Introduction – History- major achievements-Biotechnology in India.
2. Plant Tissue culture – Totipotency- definition and importance – dedifferentiation, redifferentiation and Cytodifferentiation.
3. Equipments and other requirements in tissue culture laboratory – instruments, tools, glass wares
4. Sterilization- Explants, equipments and medium
5. Culture media-MS Medium, composition and preparation
6. Inoculation – Subculture, Callus and suspension culture, meristem culture
7. Somaclonal variation- Somatic embryogenesis and organogenesis.
8. Production of haploids – pollen culture, anther culture – protoplast culture – somatic hybrids – cybrids - Synthetic seeds

MODULE-II

10hrs

1. Recombinant DNA technology:
2. General account of cloning vehicles – plasmid, bacteriophages, cosmids and phagemids. Cutting and joining of DNA molecules – restriction endonucleases, ligases – Gene library.
3. Brief account of gene transfer techniques – Direct DNA uptake by protoplast – vector method Agrobacterium mediated, physical method- electroporation- shot gun method –microinjection.

MODULE- III

14hrs

1. Methods in Biotechnology.
 - a. Isolation and purification of DNA from plant cells.
 - b. Agarose gel electrophoresis
 - c. PCR, RFLP, DNA sequencing-Sanger's method, Southern blotting, ELISA.
2. Application of biotechnology in
 - a. Medicine – edible vaccines from plants, gene therapy.

SEMESTER-VI

- b. Agriculture – *nif* genes, Genetically modified crops
- c. Industry and environment (brief account only)
3. Biosafety and ethical issues, Intellectual Property Rights (IPR)

MODULE- IV

10hrs

Microbial and Industrial Biotechnology

1. Microbes in Biotechnology.
2. Microbial culture methods of culturing, media and composition (LB and PDA)
3. Application of recombinant microbes.
4. Industrial microbiology: Production of alcohol, vinegar, bread, dairy products & single cell protein (brief account only)

MODULE- V

08hrs

Nanobiotechnology

1. Introduction-background and definition of nanotechnology
2. Nanosystems in nature - Subcellular components
3. Nanoscaled biomolecules (nucleic acids and proteins)
4. Technologies for visualization of biological structures at the nano scale-atomic force microscope
5. Nanoparticles- Quantum dots, Paramagnetic iron Oxide Crystals, Dendrimers, Carbon nanotubes.
6. Application of nanotechnology in life sciences; Biosensors

Field Study

1. Visit to a well equipped biotechnology laboratory to familiar with the use of equipments and glasswares. Petri dishes, conical flasks, culture tubes, Pasteur pipettes, forceps, scalpels, hot air oven, autoclave, platform shaker, pH meter and laminar air flow system.
2. Preparation of media, sterilization, inoculation and callus induction (demonstration only).

REFERENCES

1. Abhilasha S Muthuriya (2009) Industrial Biotechnology. Ane Books Pvt. Ltd
2. Benjamin Lewin (2004). Gene VIII Pearson Education International
3. Bharat Bhushan (2004) Hand book of nanotechnology. Springer- verlag, Berlin
4. Balasubramoniun D, CFA Bryce, K Dharmalingam, J Green and Kunthala
5. Jayaraman (2007), Concepts in Biotechnology, Universities Press
6. Channarayappa (2008), Molecular Biotechnology, Universities Press
7. Colin Ratledge (2006) Basic Biotechnology, Cambridge University Press India Pvt. Ltd
8. Gayathri M C (2015), Plant Tissue Culture: Protocols in Plant Biotechnology, Narosa Publishers Pvt Ltd
9. Gupta P. K. (2010). Elements of Biotechnology, Rastogi Publications.
10. Ignacimuthu S. J. (1996) Applied Plant Biotechnology, Tata Mc GrawHill

SEMESTER-VI

11. Janardhanan S and Vincent S (2007) Practical Biotechnology, Universities Press
12. Kesavachandran K and KV Peter (2008), Plant Biotechnology, Universities Press
13. Kumar H .D. (1983) . Molecular Biology & Biotechnology, Vikas publishing
14. Misra SP (2009) Plant Tissue Culture. Ane Books Pvt. Ltd
15. Pamela Peters (1993). Biotechnology: A guide to genetic engineering, WC Brown Publishers
16. Ramawat K. G. (2011). Plant Biotechnology, S. Chand & Company
17. Razdan M. K. (2016) An introduction to Plant Tissue Culture ,Oxford and I B H publishers
18. Reinert J. and Bajaj Y. P. S (1982). Plant cell, Tissue and Organ Culture, WC Brown publishers.
19. Smith (2008) Biotechnology, 5th Edition, Cambridge University Press India Pvt.Ltd
20. Sobti RC and Suparna S Pachauri (2009), Essentials of Biotechnology, Ane Books Pvt. Ltd
21. Subbiah Balaji (2010) Nanobiotechnology, MJP Publishers, Chennai
22. Timir Baran Jha and Biswajit Ghosh (2007), Plant Tissue Culture, Universities Press
23. Victoriano Valpuesta 2004, Fruit and Vegetable Biotechnology, CRC Press. NewYork. Ane Books Pvt. Ltd

Expected outcome

1. Students are familiarized in preparation of culture solutions, sterilization, inoculation of explants, induction of callus and morphogenesis
2. They are familiarized in biotechnological tools like RFLP, RAPD and PCR techniques
3. Use of equipments and tools in biotechnology
4. Understanding of ethical and legal issues in biotechnology and basic knowledge about IPR
5. Better understanding of nanosystems, biosensors and application of nanotechnology in biological systems

MODEL QUESTION PAPER

FIRST SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY CORE COURSE I

BO1141 : Angiosperm Anatomy, Reproductive Botany and Palynology

(2019 Admission Onwards)

Time :3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions. Each question carries 1 mark

Write short notes on

1. What is Primary wall
2. In which plant Bulliform cells are present?
3. What is Endodermis
4. Define Endarch condition
5. What is a Conjunctive tissue
6. Define Raphides
7. What are Tyloses
8. Amoeboid Tapetum
9. Comment on Anatropous ovule
10. Define Porogamy

(10 x 1 =10 marks)

SECTION B

II. Answer any Eight questions. Each question carries 2 mark

11. Write a note on Bordered pits
12. Comment on periderm and its function.
13. Mention different types of Stomata
14. Give an account of Laticifers
15. Differentiate Heart wood and sap wood
16. Give an account of Quiscent centre
17. Explain Guttation
18. Write a note on Fibers
19. Explain Polyembryony
20. Draw a neat labelled diagram of Dicot embryo with cellular divisions.
21. Write about Pollen kitt
22. Explain Kopper Kappe theory

(8 x 2 =16 marks)

SECTION C

III. Answer any six questions. Each question carries 4 marks

23. Write an account on Meristems and its classification
24. Enumerate Different types of vascular arrangement seen in Angiosperms
25. Differentiate dicot stem with that of monocot Stem.
26. With the help of diagram describe the internal structure of young anther
27. Explain different types of Endosperm formation found among Angiosperm
28. Differentiate dicot and monocot root.
29. Briefly describe the theories on apical organization of stem apex.
30. Describe the structure and function of Cambium.
31. Explain about the Non living inclusions of a cell.

(6 x4 =24 Marks)

SECTION D

IV. Answer any two questions. Each carries 15 marks.

32. What are permanent tissues? Explain different types.
33. With suitable diagrams describe normal secondary growth in Dicot stem.
34. Write an account on the process of secondary growth in Bignonia stem with diagrams
35. Explain Megasporogenesis and the *Allium* type of Embryosac development with the help of diagrams.

(2x 15=30 Marks)

MODEL QUESTION PAPER

SECOND SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : FOUNDATION COURSE II

BO1221: Methodology and Perspective in Plant Sciences

(2019 Admission Onwards)

Time: 3 Hours

Max. Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one or two sentences. Each question carries 1 mark.

Write short notes on

1. Primary data
2. Empirical knowledge
3. Range
4. Pictogram
5. Buffer
6. Median
7. Null hypothesis
8. PAGE
9. DPX
10. A killing agent

(10x1=10 marks)

SECTION B

II. Answer any 8 of the following. Each question carries 2 marks.

11. Differentiate primary and secondary source of information.
12. Comment on transparency and honesty in science.
13. Why sharing of knowledge is essential?
14. What is standard deviation?
15. Differentiate hypothesis and null hypothesis.
16. Write notes on Camera Lucida
17. Give the principle of Beer Lamberts Law.
18. What are the steps involved in maceration?
19. Write notes on double staining
20. What is range? How it is calculated?
21. Write different sources of secondary data
22. Describe parts of a table

(8x2=16 marks)

SECTION C

III. Answer any six of the following. Each question carries 4 marks.

23. What is the significance of ethics in science?
24. Differentiate inductive and deductive reasoning.
25. Briefly describe about the working of a phase contrast microscope.
26. What are the applications of SEM and TEM
27. Write brief notes on different types of stains
28. Differentiate PAGE and AGE.
29. Write brief notes on Chi square test.
30. Briefly explain the uses and applications of cryobiology.
31. Write notes on centrifugation and different types of rotors.

(6x4=24 marks)

SECTION D

IV. Write essay on any two of the following. Each question carries 15 marks.

32. Write notes on the process of killing and fixing and briefly describe the process of microtome sectioning.
33. Explain the methods of data collection and possible ways of its representation.
34. Explain different separation methods for biological molecules.
35. Discuss science as a human activity. Add notes on major revolutions in science and technology.

(2x15=30 marks)

MODEL QUESTION PAPER

THIRD SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE II

BO1341 Microbiology, Phycology, Mycology, Lichenology & Plant Pathology
(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. Name the causative organism of Citrus canker
2. Write down the male and female sex organs in the fungal partner of *Usnea*
3. What are aflatoxins?
4. Define rhizosphere
5. What is annulus
6. Define coprophilous fungi.
7. Comment on synzoospore
8. What are prions?
9. Which algae is known as stone wort?
10. Name the algal source of carageenin

(10x1 = 10 marks)

SECTION B

II. Answer any eight questions in a paragraph : Each question carries two marks

11. Write down four characteristic features of retroviruses.
12. Describe Hormogonia as a method of reproduction.
13. What is the plakea stage in *Volvox*?
14. Differentiate auxospore and autospore.
15. What are conceptacles?
16. Explain the thallus structure of *Polysiphonia*.
17. Describe the asexual reproduction in *Rhizopus*.
18. Briefly explain the asexual reproduction in yeast.
19. Explain the cleistothecium type of ascocarp in *Penicillium*.
20. Comment on ectomycorrhiza.
21. Differentiate fruticose and foliose lichen with examples.
22. Write a short note on preparation of lime sulphur

(8x2 = 16 marks)

SECTION C

III. Answer any six questions; not to exceed 120 words. Each question carries four marks.

23. Write the causative organism, symptoms and control measures of following diseases
 - a. Blast disease of paddy
 - b. Leaf mosaic disease of tapioca
24. Explain the fruiting body and sexual reproduction of *Agaricus*
25. Briefly explain the economic importance of Fungi
26. Describe the asexual and sexual reproduction in *Vaucheria*
27. Give a detailed account on classification of algae by F.E Fritsch.
28. Detail the gene transfer methods in bacteria.
29. With suitable diagrams explain the multiplication cycle of a bacteriophage.
30. Explain the thallus structure and reproduction of *Nostoc*.
31. Describe the apothecium of *Peziza* with a neat labeled diagram

(6x4 = 24 marks)

SECTION D

IV. Write an essay on any two of the following(not more than three pages)

Each question carries 15 marks.

32. With the help of neat and labelled diagrams describe the life cycle of *Puccinia*
33. What is diplontic life cycle? Explain the thallus structure, and sexual reproduction of *Sargassum*
34. Compare the nannandrous and macrandrous life cycle of *Oedogonium*
35. Describe the asexual and sexual reproductive methods in *Volvox*.

(2x15 = 30 marks)

MODEL QUESTION PAPER

FOURTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

Botany: Core course III
BO1441: Bryology, Pteridology, Gymnosperms & Palaeobotany
(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences; Each question carries one mark.

1. What is carbon dating?
2. What are amphibious plants?
3. What is calyptras?
4. Who is the father of palaeobotany?
5. What is a ligule?
6. What is gemma?
7. Name two types of rhizoids in Riccia.
8. What is ramenta?
9. Name a genus with winged pollen.
10. What is parthenocarpy?

(10x1=10 Marks)

SECTION B

II. Answer any eight of the following; Each question carries two marks

11. Write a short note on Peristome teeth.
12. Comment on Geological timescale
13. What are hydrophytic characters of *Marsilea*?
14. What is alternation of generation?
15. Enumerate the salient features of coralloid root.
16. Differentiate homospory and heterospory with examples.
17. What are the angiosperm characters of *Gnetum*?
18. Write a short note on Rhizophore of *Selaginella*.
19. Briefly explain the thallus structure of *Anthoceros*
20. Write a note on Eusporangiate ferns.
21. Explain circinate vernation with example.
22. Write a short note on transfusion tissue.

(8x2=16 marks)

SECTION C

III. Answer any six of the following; Each question carries four marks

23. What is the role of Palaeobotany in relation to the exploration of fossil fuels?
24. With a labelled diagram, explain the internal structure of *Pinus* needle.
25. Narrate the economic importance of Gymnosperms.
26. Draw the L. S. of *Cycas* ovule and discuss.
27. Enumerate the dissimilarities of bryophytes with pteridophytes.
28. Explain the structure of rachis of *Cycas*.
29. What are the economic importances of bryophytes?
30. Write down the structure of *Lepidodendron*.
31. Discuss the spore dispersal mechanism in *Pteris*.

(6x4=24 marks)

SECTION D

IV. Write an essay on any two of the following; Each question carries fifteen marks

32. Describe the stelar evolution in Pteridophytes.
33. What are fossils? Discuss various methods of fossilization.
34. Compare the sporophytes of bryophytes that you have studied.
35. Give an account of sexual reproduction in *Gnetum*. Mention the advanced characters.

(2x15=30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE IV

BO 1541- Angiosperm Morphology, Systematic Botany, Economic Botany,
Ethnobotany & Pharmacognosy

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. What is Regma ?
2. Name the inflorescence mostly found among Euphorbiaceae members.
3. What is a Legume?
4. Define Phyllotaxy.
5. What is a Floral diagram ?
6. Name the fruit of Asteraceae.
7. Define epigyny
8. Which is the source of Quinine ?
9. Morphology of cotton
10. Name two plant derived drugs used in tribal medicine

(10x1= 10 marks)

SECTION B

II. Answer any eight questions. Each question carries 2 marks. (Answer not to exceed one paragraph).

11. What is aestivation? Mention different types.
12. Differentiate between actinomorphic and zygomorphic flowers.
13. Give an account of different types of Schizocarpic fruits.
14. Write a note on Binomial nomenclature.
15. Distinguish between syngenesious and synandrous condition
16. Give the binomial and family of two dye yielding plants.
17. Explain the role of BSI in taxonomy.
18. Name an Indian ethno botanist and mention his/her major contributions
19. Narrate the salient features of Malvaceae.
20. What are the advanced features of Lamiaceae ?
21. Enlist diagnostic characters of Verbenaceae
22. 22. Explain the morphology of tendrils in Cucurbitaceae

(8x2 = 16 marks)

SECTION C

III. Answer any six questions. Each question carries 4 marks. (Answer should not exceed 120 words).

23. What is placentation? Explain different types with examples.
24. Compare the salient features of Apocynaceae and Asclepiadaceae.
25. Explain the Principles of ICBN.
26. Write an account of Herbarium techniques.
27. "Flower is a modified shoot". Comment on the statement.
28. Give an account of cereals and millets you have studied.
29. Give a brief account on APG.
30. What is Chemotaxonomy? Write down its applications
31. Explain the advanced characters of Orchidaceae?

(6x4 = 24 marks)

SECTION D

IV. Answer any two questions. Each question carries 15 marks. (Answer not to exceed three pages).

32. Describe Bentham and Hooker's system of classification. Comment on its merits and demerits.
33. Discuss the floral variations in Leguminosae
34. Discuss the advanced features of Asteraceae. Add a note on the reason for the wide distribution of Asteraceae members.
35. Give an account of different types of inflorescences you have studied.

(2x15= 30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE V

BO 1542 : Environmental Studies and Phytogeography

(2019 Admission Onwards)

Time-3 Hours

Total marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in a word or sentence, each question carries 1 mark

Write short notes on

1. A Natural Hazard
2. Halophyte
3. Food web
4. Red data book
5. Acid rain
6. Edaphic factors
7. Deforestation
8. Mitigation
9. Social forestry
10. Xerophytes

(10x1=10 marks)

SECTION B

II. Answer any 8 of the following in a paragraph. Each question carries 2 marks.

11. What are ecological pyramids?
12. Define succession
13. Briefly describe energy flow in an ecosystem
14. Write note on Ramsar sites
15. Write about impact of land slides
16. Explain 'Hot spots'
17. Enlist effects of earthquakes
18. Explain role of decomposers in an ecosystem
19. Write about Forest conservation Act
20. What is Green house effect?
21. Define Synecology
22. Explain MAB

(8x2=16 marks)

SECTION C

III. Answer any 6 of the following. Each question carries 4 marks each

23. Explain different components of an ecosystem.
24. Write about anatomical, morphological and physiological adaptations of hydrophytes
25. Explain nuclear hazards
26. Explain different stages of succession in a hydrosere
27. Give an account on adaptations of mangroves
28. Explain the process of waste water treatment
29. Explain the causes of soil degradation.
30. Explain anthropogenic effect on flood and drought
31. Explain sustainable development. Add a note on its significance

(6x4=24marks)

SECTION D

IV. Answer any two questions. Each question carries 15 marks each

32. Explain Phytogeographical regions of India.
33. Give an account of natural resources and their conservation.
34. Explain various phases of disaster management.
35. Explain different types of pollution.

(2 x 15= 30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY CORE COURSE VI

BO1543: Cell Biology, Genetics and Evolutionary Biology

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences; Each question carries one mark.

1. What is a test cross?
2. What is heredity?
3. What is telomeric chromosome?
4. Who were the rediscoverers of Mendel's work?
5. Name one characteristic each in plant and man, which show polygenic inheritance?
6. Explain genetic drift?
7. Explain aneuploidy?
8. What is SAT chromosome?
9. What is parallel evolution?
10. According to Mendel how recombinants are formed?

(10x1=10 marks)

SECTION B

II. Answer any eight of the following; Each question carries two marks

11. Briefly describe Ribosome and its major functions?
12. People with AB blood groups are called 'Universal acceptors'. Why?
13. Explain holandric genes with suitable examples?
14. Explain Lamarck's theory of evolution?
15. Why colour blindness more frequent in males than in females?
16. Mutation is the basic necessity in the origin of species. Justify the statement?
17. What are salivary gland chromosomes? Give examples?
18. What are nucleoproteins? Differentiate histones and non histones?
19. Explain Neo Darwinism?
20. Describe the role of polyploidy in evolution?
21. What do you mean by criss-cross inheritance?
22. Write notes on Triple X chromosome?

(8x2=16 marks)

SECTION C

III. Answer any six of the following; Each question carries four marks

23. Differentiate lysosome and peroxisome?
24. How can isolation of a population leads to speciation?
25. Describe the ultra structure of cell membrane?
26. Write an account on XX-XO system of sex determination?
27. Explain the genetic basis of ABO blood group in man ?
28. Describe the different types of structural chromosomal aberrations?
29. What is meant by Evolution? Give three arguments to convince a layman that evolution has takes place
30. State reasons for Mendels' success in formulating the law of heredity?
31. Are acquired characters inherited? Explain.

(6x4=24 marks)

SECTION D

IV. Write an essay on any two of the following; Each question carries fifteen marks

32. What are giant chromosomes? Briefly describe the structure and organization of any two types of giant chromosomes?
33. 'Only the first nuclear division is really meiotic and not the second division'. Explain it with the help of a diagrammatic representation of different stages of meiosis?
34. Explain various sex determination mechanisms in plants?
35. Explain linkage and crossing over? Describe its importance.

(2x15=30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

OPEN COURSE I
B01551.1 : Horticulture
(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. What is Air layering?
2. Define Hydroponics
3. What are the uses of Weedicides
4. Define Parthenocarpy
5. Write two examples for rooting hormone
6. What are Fertilizers?
7. Write common uses of Vermiculite
8. What are foliar sprays?
9. Explain NPK?
10. Define Pomology

(10x1=10 marks)

SECTION B

II. Answer any eight of the following; not to exceed one paragraph. Each question carries two marks

11. Write four principles of garden making?
12. Explain approach grafting
13. Write two examples of garden tools and its uses?
14. What are the advantages of Vermi compost
15. Explain potting mixture
16. What is carpet beds?.
17. What is sphagnum moss?
18. Write four uses indoor garden
19. What is Bouquet?
20. What is the purpose of storage of flowers?
21. Write uses and examples of fungicide
22. Briefly explain different types cutting

(8x2=16 marks)

SECTION C

III. Answer any six of the following; not to exceed 120 words; Each question carries four marks

23. Explain flower arrangements
24. What is the relevance of conservatory and green house in a garden?
25. Explain different types of layering
26. Briefly explain different types of soil
27. Describe different types of irrigation methods
28. What are the components of a garden?
29. Write an account on Bonsai
30. Explain T-budding
31. Explain Methods of Dry flower arrangements

(6x4=24 marks)

SECTION D

IV. Write an essay on any two of the following; Each question carries fifteen marks

32. Write an essay on common diseases of fruits and vegetable crops
33. Explain various types of grafting methods
34. Briefly explain different types processing and preservation of fruits and vegetables
35. Give an account of Manures and fertilizers

(2x15=30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

OPEN COURSE I

BO1551.2: Mushroom Cultivation and Marketing

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. Give the name of any two edible mushrooms.
2. What is Spawn?
3. Name any one field mushroom.
4. Give an example of medicinal mushroom.
5. What is blanching?
6. Write the scientific name of white button mushroom.
7. Name any two poisonous mushrooms.
8. What is composting?
9. What is the scientific name of milky mushroom or paddy straw mushroom?
10. What is a substrate?

(10x1=10 Marks)

SECTION B

II. Answer any eight of the following; not to exceed one paragraph. Each question carries two marks.

11. What are the advantages of mushroom cultivation?
12. Write a note on common Indian mushrooms.
13. Write a short note on field cultivation of mushroom.
14. Give an account of poisonous mushrooms.
15. Write a note on the nutritional value of mushrooms.
16. Comment on the medicinal value of mushrooms.
17. Which are the bacterial diseases that affect mushrooms?
18. Write a note on freeze drying.
19. Narrate the harvesting of mushrooms.
20. Write a note on pickling.
21. Write the systematic position of *Agaricus*.
22. Write any two binomial of *Pleurotus*.

(8x2=16 Marks)

SECTION C

III. Answer any six of the following; not to exceed 120 words. Each question carries four marks

23. Explain the Spawn preparation?
24. What are the common diseases and pests found in mushrooms?
25. Describe the various measures and methods adopted to prevent and control diseases in mushrooms.
26. Differentiate edible and poisonous mushrooms.
27. Explain pre-treatment of substrate.
28. Which are the steps involved in bed preparation.
29. Briefly explain the cultivation of white button mushroom.
30. Write a note on foreign exchange from mushroom cultivation.
31. Explain the economic importance of mushrooms.

(6x4=24 marks)

SECTION D

IV. Write an essay on any two of the following. Each question carries fifteen marks

32. Explain the distribution, structure and morphology of *Agaricus*.
33. What is the scientific name of *Oyster* mushroom? Discuss about the various steps in the cultivation of *Oyster* mushroom.
34. How mushrooms are stored? Explain the various steps in the processing of mushrooms.
35. Explain different substratum and their processing methods used in the cultivation of mushrooms.

(2x15=30 marks)

MODEL QUESTION PAPER

FIFTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

OPEN COURSE I
B01551.3 : Forestry
(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. What are Multipurpose trees?
2. What is Shellac?
3. Define Silviculture.
4. What are Tropical Forests?
5. Define Sacred plants.
6. Differentiate Homogenous and Heterogenous wood
7. Explain Avenue plantation.
8. What is NTFPs?
9. Define Seed dormancy
10. Describe Dendro chronology.

(10x1=10 Marks)

SECTION B

II. Answer any eight of the following; not to exceed one paragraph. Each question carries two marks.

11. What is gene conservation?
12. What are Gums and resins?
13. What is the sacredness of *Aeglemarmelous*?
14. Explain different types of seed dormancy
15. What do you mean by regeneration of forests?
16. Describe the types of seed dormancy.
17. Define Seed Orchard
18. Write uses of Medicinal plants
19. What are Sacred Groves?
20. Explain porous and non-porous wood
21. Write the importance of Charcoal

(8x2=16 Marks)

SECTION C

III. Answer any six of the following; not to exceed 120 words. Each question carries four marks.

22. Briefly explain types of seed dormancy
23. Point out importance of Evergreen and deciduous forest in forest ecosystem
24. What is the relevance of wood anatomical studies in Kerala?
25. Give an account of Timber Forest Products
26. Write about Silviculture of *Azadirachta indica*
27. Explain coppice and conservation systems
28. What are the uses of canes and bamboos?
29. Write an account of Trees in Sacred groove
30. Write about Lac collection and marketing
31. Explain methods to overcome seed Dormancy

(6x4=24 Marks)

SECTION D

IV. Write an essay on any two of the following. Each question carries fifteen marks.

32. Write an essay on Silviculture methods
33. Explain various types of Forest in Kerala
34. Briefly explain different types of wood
35. Explain Indian forest act 1927 and their amendment

(2x15=30 Marks)

MODEL QUESTION PAPER

SIXTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE IX

BO 1641 - Plant Physiology and Biochemistry

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark

1. What is Kranz anatomy?
2. What are epimers?
3. Why cytochrome a_3 is called terminal oxidase?
4. Represent the overall equation of photosynthesis.
5. What are isoenzymes?
6. What is meant by thigmotropic movement?
7. What is tonicity?
8. Define red drop.
9. What is incipient plasmolysis?
10. How the stomata of CAM plants differ from that of C3 plants?

(10 x 1 = 10 marks)

SECTION B

II. Answer any 8 questions. Each question carries 2 marks.

11. Give an account on raw materials of photosynthesis.
12. Briefly describe the physiological role of Abscisic acid in plants.
13. Differentiate apoplast and symplast.
14. How root pressure influences water movement in plants?
15. What are derived lipids? Give examples
16. Explain the physiology of senescence.
17. Discuss the amphoteric nature of aminoacids
18. Briefly describe the relation between OP, TP and WP in plants.
19. Comment on hydroponics.
20. Differentiate absorption spectrum and action spectrum.
21. Why fluorescent radiation has higher wavelength than phosphorescent radiation?
22. Point out the differences between photosynthesis in bacteria and higher plants.

(8 x 2 = 16 marks)

SECTION C

III. Answer any six of the following. Each question carries 4 marks

23. Describe the vital and physical theories explaining ascent of sap.
24. How the principle of Limiting factors apply in photosynthesis?
25. Give an account on coenzymes and co-factors.
26. Explain beta oxidation of fatty acids.
27. Describe the classification and function of secondary plant products.
28. Explain the mechanisms of mineral absorption in plants.
29. How anaerobic respiration differs from aerobic in terms of ATP yield?
30. Discuss the mechanism of photorespiration. Enumerate its advantages and disadvantages
31. Explain biological nitrogen fixation.

(6 x 4 = 24 marks)

SECTION D

IV. Answer any 2 questions. Each question carries 15 marks.

32. Discuss the mechanisms and theories on translocation of organic substances in plants.
33. Explain dark reaction and compare the photosynthetic mechanisms in C₃ and CAM plants.
34. Discuss various types of movements exhibited by plants.
35. What are carbohydrates? How are they classified? Mention its function

(2x15 = 30 marks)

MODEL QUESTION PAPER

SIXTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE X

BO-1642-Molecular Biology, General Informatics and Bioinformatics

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark

1. Define cistron.
2. How is wet lab different from a web lab?
3. Expand NICNET
4. Name two operating systems used in computers.
5. Mention role of helicases.
6. Explain PIR
7. What are okazaki fragments?
8. Name two nucleic acid databases.
9. What are introns?
10. Write about importance of Reverse transcriptase

(10x1=10 marks)

SECTION B

II. Answer any eight of the following: Each question carries two marks

11. Give any four differences between DNA and RNA.
12. What is Kornberg enzyme.
13. What is plagiarism?
14. What are stop codons? Give their sequence
15. What are replisomes?
16. Describe the health issues faced by regular users of computer.
17. What is SWISS-PROT? Mention any two important features.
18. What you mean by the statement “the genetic code is degenerate”?
19. Write a note on the main objectives of INFLIBNET.
20. Differentiate between Global and Local alignment
21. What are the parts of a typical rooted phylogenetic tree?
22. How CLUSTAL becomes an indispensable tool in Bioinformatics?

(8x2= 16 marks)

SECTION C

III. Answer any six of the following: Each question carries four marks

23. Give an account on sequence analysis and alignment in bioinformatics.
24. Write the salient features of Watson and Crick double helical model of DNA.
25. Comment on intellectual property rights.
26. Give an account on operon concept with reference to *lac* operon.
27. What is a phylogram? Describe the various tools used in phylogenetic analysis.
28. Give an account of the different types of RNA
29. Practicing good cyber ethics is a present day necessity. Comment.
30. Describe the experiment of Meselson and Stahl
31. How is Rasmol used in molecular visualization?

(6x4=24 marks)

SECTION D

IV. Write essay on any two of the following. Each question carries fifteen marks

32. What are the advantages of molecular phylogenetic analysis? Explain the technique
33. Describe the various experiments conducted by scientists to prove that DNA is the genetic material in living organisms.
34. Give a detailed account on different biological databases.
35. What is central dogma in molecular biology? Elucidate the process by which genetic information is transferred from DNA to RNA in a prokaryotic cell.

(2x15 = 30 marks)

MODEL QUESTION PAPER

SIXTH SEMESTER B. SC. DEGREE (CBCSS) EXAMINATION

BOTANY : CORE COURSE XI

BO1643: Horticulture, Plant Breeding & Research Methodology

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark.

1. What is turf culture?
2. What are mutagens?
3. Define Pomology.
4. What is pure research?
5. What is urkund?
6. What is floriculture?
7. What is NPK?
8. What is heterosis?
9. What is the application of peat moss?
10. Define acclimatization.

(10x1= 10 marks)

SECTION B

II. Answer any eight of the following; Each question carries two marks.

11. Give an account on biofertilizers used in horticulture.
12. Write a short note on quarantine.
13. What is foliar spray?
14. Explain the advantages of organic manures.
15. Differentiate horizontal resistance from vertical resistance.
16. What is null hypothesis?
17. Describe backcross method.
18. Explain air layering.
19. Make a short note on author citation.
20. What is bonsai?
21. Write short notes on plagiarism.
22. Mention the types of grafting.

(8x 2= 16 Marks)

SECTION C

III. Answer any six of the following; Each question carries four marks.

23. Differentiate pureline selection from mass selection.
24. Describe polyploidy breeding.
25. Describe the method of vermi-composting.
26. Make an account on project report presentation.
27. Explain inter-varietal hybridization.
28. Describe the procedure involved in data collection.
29. Explain the steps to be taken for weed control.
30. Write an account on types of flower arrangement.
31. Explain indoor gardening.

(6x4=24 marks)

SECTION D

IV. Answer any two of the following; Each question carries fifteen marks.

32. Explain various grafting techniques in plant propagation.
33. Method and application of mutation breeding.
34. Write a detailed account on various garden implements and their uses.
35. Explain scientific method in biological research.

(2x15 = 30 Marks)

MODEL QUESTION PAPER

SIXTH SEMESTER B SC DEGREE (CBCSS) EXAMINATION

OPEN COURSE –II – ELECTIVE

BO 1651: Biotechnology and Nano Biotechnology

(2019 Admission Onwards)

Time. 3 Hrs

Total Marks: 80

(Draw diagrams wherever necessary)

SECTION A

I. Answer all questions in one word or two sentences. Each question carries one mark

1. Define totipotency
2. Define callus
3. What is meant by surface sterilization?
4. What are cybrids
5. What are edible vaccines
6. Define cosmids
7. Define quantum dots
8. What are single cell proteins?
9. Define gene library
10. Define patent

(10x1=10 marks)

SECTION B

II. Answer any eight of the following. Each question carries two marks

11. Differentiate between redifferentiation and redifferentiation
12. What are synthetic seed? Mention the uses
13. What are nif genes? Mention the features of nif genes
14. What is meant by somaclonal variation?
15. What are genetically modified crops? Mention any two GMC you have studied.
16. What is meristem culture? Mention its advantages.
17. Define plant growth regulators? Give examples
18. What is meant by Cryo preservation?
19. Give a brief account on Somatic embryogenesis
20. What is protoplast culture?
21. What is Ti Plasmid?
22. Explain the composition and preparation of PDA medium

(8x2=16 marks)

SECTION –C

III. Answer any six of the following; Each question carries four marks

23. Give an account on Somatic hybridization.
24. Discuss the classification of Nanoparticles in detail.
25. Give an account on suspension culture and its applications
26. Give a brief account on various gene transfer mechanisms in plants
27. Write about Southern blotting
28. Write a note on recombinant microbes and its applications.
29. Define nanoparticle. Explain the structure and properties of nanoparticles.
30. Define sterilization. Give an account on various sterilization techniques used in plant tissue culture.
31. Explain the technique of electrophoresis.

(6x4=24 marks)

SECTION -D

IV. Write essay on any two of the following; Each question carries fifteen marks

32. Explain the composition and preparation of MS medium
33. Define Protoplast culture. Explain the steps involved in protoplast culture and its application.
34. Briefly explain Recombinant DNA Technology
35. What is PCR? Explain the basic steps and its application

(2x15=30 marks)

MODEL QUESTION PAPER

FIRST DEGREE PROGRAMME IN BOTANY (CBCSS) PRACTICAL EXAMINATION

Botany Core (Practical I) VII-Course Code: BO1544
Angiosperm Anatomy, Reproductive Botany, Palynology,
Methodology and Perspectives in Plant Science
(2019 Admission Onwards)

Time: 3 Hours

Max. Marks: 80

1. Make suitable micropreparation of **A**, identify giving reasons and describe its structure with the help of labelled diagrams. Leave the preparation for valuation. (Preparation-3, Labelled diagram-3, Reasons-3, Identification-1) **10 Marks**
2. Construct a frequency table (Discrete/Continuous) from the given data **B** **4 Marks**
3. Construct histogram/Bar diagram from the data **C**. **4 Marks**
4. Workout the problem **D**
(Calculation- 3, Result-1) **4 Marks**
5. Identify the instrument **E**. Comment on its working principle.
(Identification-1, Aim-1, Working Principle-2) **4 Marks**
6. Identify the separation technique **F**. Comment on its procedure.
(Identification-1, Aim-1, Procedure-2) **4 Marks**
7. Comment on **G**
(Major group-1, Notes-2) **3 Marks**
8. Write critical notes on **H**
(Identification -1, Notes-2) **3 Marks**
9. Make suitable micropreparations of **I**, identify giving reasons and describe its structure with the help of labelled diagrams. Leave the preparation for valuation. (Preparation-3, Labelled diagram-3, Reasons-3, Identification-1) **10 Marks**
10. Identify the type of stomata in specimen **J**
(Identification-1, Labelled Diagram-1, Reasons-1) **3 Marks**
11. Identify and describe the type of cellular inclusions in specimen **K**
(Identification-1, Description-2, Labelled Diagram-1) **4 Marks**
12. Identify **L** and draw a neat labelled diagram
(Identification-1, Labelled Diagram-3) **4 Marks**
13. Comment on **M**
(Identification-1, Notes-2) **3 Marks**
- Record (Content-15, Neatness-5) 20 Marks**

MODEL QUESTION PAPER

KEY TO SPECIMENS

- A. Primary/Normal Secondary Root/Stem
- B. Numerical data
- C. Frequency table data
- D. Frequency table/Numerical Data
- E. Instruments from Biophysics
- F. Biophysics- Chromatography
- G. Fixatives/ Stains/ Mounting media
- H. Dissection/Compound microscope/Microtome/Camera lucida/Micrometer mentioned in the syllabus
- I. Stem with anomalous secondary structure (*Bignonia, Boerhaavia, Dracaena*)
- J. Stomatal type-
- K. Cell inclusions mentioned in the syllabus
- L. Anther T. S /Dicot embryo L. S/ Monocot embryo L.S. Embryosac (*Polygonum* type) Ovule (Orthotropous/ Anatropus) (*Permanent slides /diagrams*)
- M. Pollen Aperture type mentioned in the syllabus

MODEL QUESTION PAPER

FIRST DEGREE PROGRAMME IN BOTANY (CBCSS) PRACTICAL EXAMINATION

Botany Core (Practical II) VIII-Course Code: BO1545
Microbiology, Phycology, Mycology, Lichenology & Plant Pathology
Bryology, Pteridology, Gymnosperms & Paleobotany
(2019 Admission Onwards)

Time: 3 Hours

Max. Marks: 80

1. Make suitable micro preparations to bring out the structure of **A, B, C** and **D**. Draw a cellular diagram of each and label the parts. Identify giving reasons and leave the preparation for valuation.
(Preparation-2, Labelled diagram-2, Reasons, 2, Identification-1)
4 x 7 = 28 Marks
 2. Identify any two algal specimens from the mixture **E**, giving reasons
(Diagram-1, Reasons-1, Identification-1)
2 x 3 = 6 Marks
 3. Observe **F** under microscope, identify and write down its procedure.
(Procedure-3, Identification -1)
4 Marks
 4. Identify the disease in plant specimen **G** and give the name of the causative organism along with important symptoms associated with it.
(Disease-1, Pathogen-1, Symptoms-2)
4 Marks
 5. Spot at sight **H, I, J, K, L** and **M**
(Genus name-1, Part of the plant-1, Major group-1)
6 x 3 = 18 Marks
- Record (Content-15, Neatness-5) 20 Marks**

KEY TO SPECIMEN

- A. Fungus mentioned in the syllabus
- B. Bryophyte mentioned in the syllabus
- C. Pteridophyte mentioned in the syllabus
- D. Gymnosperm mentioned in the syllabus
- E. Algal Mixture (Mixture of different algae (at least four members) mentioned in the syllabus.
- F. Bacterial Smear (Gram positive/ Gram negative)
- G. Plant disease mentioned in the syllabus
- H. Alga (Macroscopic)
- I. Fossil form mentioned in the syllabus(Permanent slide /Photograph)
- J. Fungus/Lichen
- K. Bryophyte
- L. Pteridophyte
- M. Gymnosperm

MODEL QUESTION PAPER

FIRST DEGREE PROGRAMME IN BOTANY (CBCSS) PRACTICAL EXAMINATION

Botany Core (Practical III)XII-Course Code: BO1644
Angiosperm Morphology, Systematic Botany, Economic Botany, Ethnobotany,
Pharmacognosy and Environmental Studies
(2019 Admission Onwards)

Time: 3 Hours

Max. Marks: 80

1. Identify and write notes on **A** with a labelled diagram.
(Notes-2, Labelled diagram-1) **(3 Marks)**
2. Comment on **B**
(Notes 2, Diagram-1) **(3 Marks)**
3. Describe **C** in technical terms. Draw a labelled sketch of the L. S. of the flower **C**. Construct a floral diagram and write the floral formula.
(Description-2, Labelled sketch-2, Floral diagram-1, Floral formula-1) **(6 Marks)**
4. Refer **D** and **E** to their respective families, pointing out the class, subclass and series with reasons. (Reasons upto series-2, Description of plant in technical terms- 2, Family characters-2, Identification of family-1) **(7 x 2 = 14 Marks)**
5. Write the generic name and family of **F** and **G**
(Generic name-1, Family-1) **(2 x 2 = 4 Marks)**
6. Spot at sight **H** and **I**
(Binomial-1, Family-1, Morphology of the useful part-1) **(3 x2 = 6 Marks)**
7. Identify and write critical notes on **J**
(Binomial -1, Family-1, Notes-2) **(4 Marks)**
8. Make a suitable micropreparation of **K**. Write the anatomical adaptations and identify the ecological group.
(Preparation-1, Adaptations-2, Ecological group-1) **(4 Marks)**
9. Identify the ecological group **L** and write its morphological adaptations.
(Ecological group-1, Adaptations-2) **(3 Marks)**
10. Identify and write notes on **M**
(Identification-1, Notes-2) **(3 Marks)**
- ☒ Herbarium submission -7; Field Note & Tour Report-3 **(10 Marks)**
- ☒ Record (Content-15 Marks. Neatness-5 Marks) **(20 Marks)**

KEY TO SPECIMEN

- A. Inflorescence (*Mentioned in the syllabus*)
- B. Fruits (*Mentioned in the syllabus*)
- C. Systematic Botany – Flowering twigs (along with floral buds) from **Polypetalae / Gamopetalae**
- D. Any Dicot family mentioned in the syllabus (Flowering twigs from **Polypetalae/ Monochlamydeae**)
- E. Any Dicot family mentioned in the syllabus (Flowering twigs from **Gamopetalae**)
- F & G. Herbarium sheets
- H & I. Economic Botany (*Mentioned in the syllabus*)
- J. Ethnobotany (*Plants mentioned in the syllabus only*)
- K & L. Ecological groups mentioned in the syllabus
- M. Ecosystems (*Photographs, Pictures or Diagrams of ecosystems mentioned in the syllabus*)

MODEL QUESTION PAPER

FIRST DEGREE PROGRAMME IN BOTANY (CBCSS) PRACTICAL EXAMINATION

BOTANY CORE (PRACTICAL IV) XIII-COURSE CODE: BO1645

**Cell Biology ,Genetics, Plant Physiology , Biochemistry,Molecular Biology, Plant
Breeding and Horticulture**

(2019 Admission Onwards)

Time: 3 Hours

Max. Marks: 80

1. Make acetocarmine squash preparation of material **A**. Identify metaphase and anaphase stages of cell division and draw labelled sketches of identified stages.
[Preparation-4, Identification-4 (2x2), labelled diagram- 4 (2x2)] **12 Marks**
 2. Work out Problems **B, C, D**
(B-3 marks, C-4 Marks, D-5 Marks) **12 Marks**
 3. With the help of a labelled diagram, explain the working of the experiment **E**.
(Aim-1, Labelled diagram-2, Working-3, Inference-1) **7 Marks**
 4. Comment on **F** with a labelled diagram
(Aim-1, Comment-3, Labelled diagram-2) **6 Marks**
 5. Detect the presence of proteins/reducing sugar/non reducing sugar/starch in **G**.
Write down the procedure.
(Conduct of experiment-1, Procedure-3, Identification-1) **5 Marks**
 6. Workout Problem **H** **4 Marks**
 7. Write down the procedure and demonstrate emasculation using material **I**. Submit for valuation.
(Demonstration-3, Procedure-2) **5 Marks**
 8. Write down the procedure and demonstrate T-Budding/Air layering/Grafting (Wedge/Whip) using **J**. Submit for valuation.
(Procedure-3, Demonstration-3) **6 Marks**
 9. Identify and write notes on **K**.
(Identification-1, Notes-2) **3 Marks**
- Record (Content- 15, Neatness-5) **20 Marks****

KEY TO SPECIMENS

- A. Onion Root tips
- B. Genetic Problem (*Monohybrid/ Incomplete Dominance*)
- C. Genetic problem (*Dihybrid*)
- D. Genetic Problem (*Interaction of Genes*)
- E. Physiology experiments
- F. Physiology experiments
- G. Biochemistry (proteins/ reducing sugar/non reducing sugar/starch)
- H. Molecular biology (Problems on Chargaff's rule)
- I. Plant Breeding: Inflorescence coming under *Fabaceae/Caesalpinaceae*
- J. Horticulture
- K. Garden tools mentioned in the syllabus

BO1646: PROJECT

- ✎ The project work/ Dissertation can be carried out individually or group of students. Project report, duly attested by the Supervising teacher and certified by Head of the Department, has to be submitted on the day of Core Practical III examination. Viva voce has to be conducted individually.

Project Report- 80 Marks Viva voce - 20 marks Total – 100 Marks